Room for Improvement: Immunizations for Patients With Monoclonal B-Cell Lymphocytosis or Chronic Lymphocytic Leukemia

Jennifer A. Whitaker, MD, MS, Tait D. Shanafelt, MD, Gregory A. Poland, MD, and Neil E. Kay, MD

The authors are affiliated with the Mayo College of Medicine in Rochester, Minnesota. Dr Whitaker is an assistant professor of medicine in the Department of Medicine in the Divisions of General Internal Medicine and Infectious Diseases. Dr Shanafelt and Dr Kay are professors of medicine in the Department of Medicine in the Division of Hematology. Dr Poland is a professor of medicine in the Department of Medicine in the Division of General Internal Medicine.

Address correspondence to: Jennifer A. Whitaker, MD, MS Assistant Professor of Medicine Mayo College of Medicine Department of Medicine Divisions of General Internal Medicine and Infectious Diseases 200 First Street SW Rochester, MN 55905 Phone: 507-284-2117 E-mail: whitaker.jennifer@mayo.edu

Keywords

Immunization, vaccine, chronic lymphocytic leukemia (CLL), monoclonal B-cell lymphocytosis (MBL) **Abstract:** Infection is the cause of death in 30% to 50% of patients with chronic lymphocytic leukemia (CLL). A major strategy to decrease infection risk is vaccination. However, vaccine response rates in patients with CLL are typically insufficient. Recent studies have demonstrated that individuals with clinical monoclonal B-cell lymphocytosis (MBL), the precursor to CLL, also have an increased risk of infection and thus could benefit from vaccines. However, there are no data on vaccine responses in the MBL population. This article reviews the immunodeficiency of CLL and MBL, discusses the recommended vaccines and data on vaccine immunogenicity in patients with CLL, and outlines the need to develop more effective vaccine strategies in this population of patients at high risk for infection.

Introduction

Vaccination has been lauded as the most effective method to prevent infectious diseases. However, some populations do not reap the same protective benefits from vaccines as others. Predominant among these populations are patients with hematologic malignancies, who are particularly vulnerable to infections and in need of effective strategies to curb infection risk. Chronic lymphocytic leukemia (CLL) is one of the most common hematologic malignancies, comprising 11% of all hematologic malignancies, with an annual incidence in the United States approaching 15,000.1 Infection is a major cause of mortality in patients with CLL, accounting for 30% to 50% of all deaths.² Strategies to reduce infections in patients with CLL have relied upon the administration of intravenous immunoglobulin (Ig), antimicrobial prophylaxis, treatment of the underlying disease, and vaccines. Despite the fact that certain vaccines are routinely recommended for patients with CLL (eg, influenza and pneumococcal vaccines), previous studies have shown low rates of immunogenicity in this population. Herein, we discuss the recommended vaccines for patients with CLL, comprehensively review the literature regarding the immunogenicity of these vaccines, provide an update on the currently known mechanisms of decreased vaccine immunogenicity, and propose solutions and the research needed to

provide more effective vaccine approaches for the CLL population. Furthermore, there are no data on the vaccine responses of individuals in the precursor state to CLL, which is monoclonal B-cell lymphocytosis (MBL). This population likely comprises 3% to 12% of adults past the age of 40 years.³⁻⁷ Given the high prevalence of MBL, and the evidence that at least some individuals with MBL are at increased risk for infection,⁸ we also discuss the need for research on vaccine efficacy in the MBL population.

Infection Risk in Monoclonal B-Cell Lymphocytosis and Chronic Lymphocytic Leukemia

Susceptibility to infection in CLL results from a complex immunodeficiency state that includes defects in innate, humoral, and cell-mediated immunity, which are present in most patients even before exposure to chemotherapy. MBL is characterized by the presence of clonal B cells in peripheral blood at a concentration below 5×10^9 /L in the absence of lymphadenopathy, cytopenias, or autoimmune disease.^{3,9,10} Although only a fraction of cases of MBL are clinically recognized (ie, patients with so-called high-count or clinical MBL), MBL is present in approximately 3% to 12% of the general population older than 40 years of age evaluated by sensitive testing.³⁻⁷ A recent case-control study of community-based individuals with high-count MBL and CLL seen at Mayo Clinic demonstrated that individuals with clinical MBL are at a 3-fold higher risk of hospitalization for infection compared with a control population after adjustments for age, sex, and comorbidities.⁸ Indeed, at 4 years after diagnosis, individuals with clinical MBL were approximately 4 times more likely to be hospitalized for infection than to require treatment for progressive CLL. The risk of hospitalization for infection among patients with MBL is similar to that among patients with CLL, both with a hazard ratio of approximately 3 relative to age-, sex-, and comorbiditymatched controls.8 The extent and complete nature of the immunodeficiency in MBL have yet to be elucidated, but the immunodeficiency is thought likely to be due to the same mechanisms that cause immunodeficiency in CLL.

Only one study has characterized the most prevalent infections in the MBL population⁸; however, the literature regarding infectious complications in the CLL population is more extensive. The case-control study of infectious complications among patients with MBL or CLL revealed that infections with gram-positive organisms are more common among patients with CLL or MBL than in controls. *Staphylococcus aureus* was the primary pathogen identified from any culture site, with *Streptococcus pneumoniae* and *Pseudomonas* species next most common.⁸ Other studies have demonstrated that *S pneumoniae* and

S aureus are common pathogens in patients with CLL, and that the respiratory tract and bloodstream tend to be the most frequent sites of infection.^{2,11,12} Although infections with these common bacterial agents often occur in patients who have untreated CLL, infections associated with immunosuppression (eg, invasive aspergillosis, nocardiosis, cytomegalovirus infection, Pneumocystis jirovecii infection) typically occur in patients who have received T-cell-depleting treatments for CLL (eg, purine analogues, alemtuzumab).8,12 Recurrent viral infections, particularly those due to herpes simplex virus and varicella-zoster virus, are also more common after treatment,13 whereas endemic mycoses tend to be associated with prolonged and profound neutropenia in patients with advanced disease.¹⁴ To our knowledge, no studies have compared the rates and severity of respiratory infections caused by viral pathogens, such as influenza virus, among patients who have MBL or CLL with those in a control population.

Immunodeficiency That May Be Related to Impaired Vaccine Responses in Chronic Lymphocytic Leukemia

In the past, the immunodeficiency of CLL was thought to be due primarily to hypogammaglobulinemia, which affects approximately 25% of patients with CLL.¹⁵ Patients with hypogammaglobulinemia tend to have an elevated risk for infections caused by bacteria, particularly encapsulated organisms.¹¹ There may be a particular infection risk related to Ig class deficiency; both IgG3 and IgG4 subclass deficiencies are associated with an increased risk of infection in patients with CLL.¹⁶ IgG3 deficiency correlates with increased risk for herpes infection, and IgG4 deficiency correlates with respiratory tract infections.¹⁷ IgA deficiency and defects in mucosal immune function may also be associated with an increased frequency of upper respiratory tract infections.¹⁸ The pathogenesis of hypogammaglobulinemia is not completely elucidated but is at least related to a progressive decline in the numbers of functional B cells¹³ and also to the suppression of residual normal B-cell function.^{13,19} With fewer functional B cells, vaccines whose protection depends on adaptive humoral immunity will obviously be less effective in generating protective antibody responses for a given patient with CLL. Most current vaccines mediate protection mainly through the induction of highly specific serum IgG antibodies,²⁰ although a few live viral vaccines induce serum IgA and secretory IgA responses (ie, intranasal influenza virus and rotavirus vaccines, oral poliovirus vaccines).20

Although hypogammaglobulinemia clearly plays a role in infection risk,^{16,21-23} newer studies have demonstrated that patients with CLL who have normal serum Ig

levels still remain at increased risk of infection.8 In addition to decreased functional B-cell numbers and Ig levels, T-cell dysfunction occurs, even in the absence of chemotherapy.²⁴ Increases in T-regulatory cells,²⁵ decreases in T-helper activity, reversal of the CD4/CD8 ratio,²⁶ defects in natural killer cells,²⁷ increases in functionally naive T cells,²⁸ and impaired T-cell immunologic synapse²⁹ have all been reported. This T-cell dysfunction places patients at risk for infection with viral pathogens, as well as other pathogens, such as fungal and mycobacterial organisms. In terms of vaccination, T-cell dysfunction is of primary importance for vaccines that rely on T-cell-dependent B-cell responses. There is evidence that T-cell responses may contribute to the protection conferred by some live attenuated viral vaccines and possibly the acellular pertussis vaccine.²⁰ T-cell dysfunction likely plays a major role in the decreased responses to vaccines in CLL because patients with CLL have significant quantitative and qualitative abnormalities in T cells.

As noted, most current vaccines mediate protection through the induction of highly specific serum antibodies via B cells. There are 2 mechanisms by which an efficient and appropriate antibody response is accomplished. One mechanism is a T-independent response by which capsular polysaccharides elicit B-cell responses in the marginal zone and extrafollicular areas of the spleen and lymph nodes. Pure polysaccharide vaccines (Table 1) rely on a T-independent pathway to induce a humoral response. The other mechanism is a T-dependent pathway by which foreign peptide antigens are presented to the immune system and recruit antigen-specific CD4+ T-helper cells. Vaccines that rely on a T-dependent pathway to induce a humoral response include the following: glycoconjugate vaccines (in which a bacterial polysaccharide is conjugated to a protein carrier), toxoid vaccines, protein vaccines, and inactivated, live, or inactivated live viral vaccines (see Table 1).²⁰ For these vaccines, the combination of both B-cell dysfunction and T-cell dysfunction will affect vaccine responses.

Other mechanisms of immunodeficiency in CLL include neutropenia (as a side effect of chemotherapy or due to autoimmune granulocytopenia), abnormal complement activity, age, effects of chemotherapy, and effects of the corticosteroids used to treat CLL or associated autoimmune cytopenias. Immunosenescence related to age and impaired vaccine responses in older adults are multifactorial processes and topics of intense research.³⁰⁻³² Cytotoxic chemotherapy has multiple and various adverse effects on immune responses to vaccines.^{33,34} In addition, corticosteroids, which are often given to patients with CLL in relation to treatment approaches or autoimmune complications, can dampen vaccine responses.³⁵⁻³⁷ In general, the Advisory Committee on Immunization Practices (ACIP) immunization guidelines^{38,39} and the Infectious

Diseases Society of America (IDSA) immunization guidelines for immunocompromised patients⁴⁰ consider prednisone at a daily dose of 20 mg or higher (or its equivalent for other corticosteroids) for 2 weeks or longer as "highdose" corticosteroid use, and immunosuppressive.

Vaccine Recommendations and Immunogenicity Studies in Chronic Lymphocytic Leukemia

MBL is considered a precursor state rather than a hematologic malignancy and is thus not currently classified as an immunocompromised state in the ACIP^{38,39} or IDSA immunization guidelines.⁴⁰ Nonetheless, more recent data suggest that at least the subset of patients with clinical MBL are at high risk for infection and are probably best approached in a manner similar to that used for individuals with CLL. Patients with CLL are considered immunocompromised in the ACIP and IDSA immunization guidelines.

Influenza Vaccines

Annual influenza immunization is recommended for all persons in the United States aged 6 months or older. Accordingly, all patients with MBL or CLL should receive influenza vaccine annually. In the United States and many other parts of the world, high-dose influenza vaccines, with 4 times the amount of hemagglutinin compared with standard-dose influenza vaccines, have been licensed for adults 65 years of age or older. The ACIP and IDSA guidelines do not specifically endorse high-dose influenza vaccines over standard-dose influenza vaccines for adults aged 65 years or older who are immunocompetent or immunocompromised.^{38,40} However, several studies have demonstrated increased immunogenicity for the highdose influenza vaccine in immunocompetent adults aged 65 years or older.^{41,42} Results of a large-scale, 2-season confirmatory efficacy trial involving more than 30,000 participants 65 years of age or older are expected to be released in 2014 by Sanofi Pasteur. Preliminary reports have indicated that Fluzone High-Dose (Sanofi Pasteur) has clinical efficacy superior to that of the standard dose of Fluzone (Sanofi Pasteur) in preventing influenza.⁴³

In the United States, only aluminum adjuvant vaccines are approved by the US Food and Drug Administration (FDA). However, in Europe and many other parts of the world, oil-in-water adjuvants, such as AS03 (Adjuvant System 03, GlaxoSmithKline) and MF59 (Novartis), have been approved for influenza vaccination. Studies have shown increased immunogenicity, and in some cases increased efficacy, with these adjuvant vaccines in comparison with standard influenza vaccines in elderly persons aged 65 years or older⁴⁴⁻⁵⁰ and in patients with hematologic malignancies.^{51,52} In the future, updated guidelines

Vaccine	Vaccine Type							
	Protein	Polysaccharide	Polysaccharide/Protein Conjugate	Toxoid	Killed Viral	Killed Bacterial	Live Attenuated Viral	Live Attenuated Bacterial
Advised to give in chronic lymphocytic leukemia								
Influenza	 ✓ 							
Pneumococcal polysaccharide		~						
Pneumococcal conjugate			~					
Tetanus toxoid (Td)				~				
Pertussis, acellular (Tdap)	 ✓ 							
Diphtheria toxoid (Td or Tdap)				~				
Permissible to give in chronic lymphocyt	ic leukemia*							
Pertussis, whole cell						~		
Hepatitis A					~			
Hepatitis B	 ✓ 							
Hib polysaccharide		~						
Hib conjugate			~					
Japanese encephalitis (inactivated)					~			
Meningococcal polysaccharide		~						
Meningococcal conjugate			~					
Polio, Salk					~			
Rabies					~			
Typhoid polysaccharide (injectable)		~						
Contraindicated in chronic lymphocytic	leukemia					1	1	
Influenza, intranasal							~	
Measles							~	
Mumps							~	
Polio, Sabin							~	
Rubella							~	
Tuberculosis (BCG)								~
Oral typhoid								~
Varicella (chickenpox)							~	
Varicella (zoster)							~	
Yellow fever							~	

Table 1. Recommended and Contraindicated Vaccines for Patients With Chronic Lymphocytic Leukemia

Td, combined tetanus and diphtheria vaccine; Tdap, combined tetanus, diphtheria, and acellular pertussis vaccine; Hib, *Haemophilus influenzae* type b; BCG, bacille Calmette-Guérin.

* Indication for these vaccines depends on the patient's previous vaccination history, occupation, travel, and medical comorbidities.^{38,39}

may recommend the use of high-dose influenza vaccines for adults aged 65 years or older. Once additional wellplanned clinical trial studies have evaluated the safety and immunogenicity of the high-dose influenza vaccines in immunocompromised populations, we anticipate updated recommendations for these groups.

There are limited data regarding influenza vaccine immunogenicity in patients with CLL (Table 2). An older study, by Marotta and colleagues, evaluated the response to 2 doses of an inactivated whole-virus vaccine administered 31 days apart.53 This study noted an association between vaccine response (defined as a hemagglutinin inhibition assay [HI] titer >1:10) and the absolute numbers of CD4+/ CD45RA+ naive T cells and CD5- B cells.53 It is difficult to extrapolate vaccine immunogenicity data from this study because the authors did not use a standard immunogenicity definition of seroconversion (a prevaccination HI titer <1:10 and a postvaccination HI titer \geq 1:40, or a prevaccination HI titer≥1:10 and a minimum 4-fold rise in postvaccination HI antibody titer) or of seroprotection (HI titer ≥1:40).⁵⁴ Another older study, by Bucalossi and colleagues, used the same definition of vaccine response as did the study of Marotta and colleagues. The investigators administered 2 doses of an inactivated influenza vaccine 31 days apart and noted an association between vaccine response and disease stage and between vaccine response and serum Ig levels.⁵⁵ It is difficult to draw meaningful inferences from these studies because they did not employ standard definitions of vaccine response.

The only study that defines influenza vaccine response by seroconversion (eg, 4-fold rise in postvaccination HI antibody titer to indicate seroconversion) is the 2001 study by van der Velden and colleagues. In this study, 20 patients with CLL at various Rai stages received 2 doses of an inactivated subunit vaccine 21 days apart. Seroconversion after 1 dose of vaccine was only 5% for influenza A strains H1N1 and H3N1 and was 15% for the influenza B strain in the vaccine. Seroconversion after 2 doses of vaccine was 15% for the 2 influenza A strains and 30% for the influenza B strain; however, vaccine response rates did not increase significantly after the booster dose.⁵⁶ Higher serum Ig levels correlated with protective HI titers for influenza B strains. Despite its small sample size, this is the only study in the literature providing information on the immunogenicity of the current influenza inactivated subunit vaccines. The results suggest that influenza vaccine seroconversion after a single dose in patients with CLL is approximately 5% for influenza A strains and 15% for influenza B strains. These responses are, of course, unacceptably low for a CLL patient cohort. Influenza vaccine serum antibody responses depend on age, preexisting antibody levels, and underlying medical conditions. A quantitative review of 31 influenza vaccine antibody response studies for standard-dose influenza vaccine found influenza seroconversion rates of roughly 60% for influenza A and B strains and seroprotection rates of roughly 80% for adults younger than 58 years. For those 58 years of age or older, the influenza seroconversion rates were 35% to 51% for influenza A and B, and the

seroprotection rates were roughly 70%.⁵⁷ There are no data on responses to high-dose influenza vaccines in the CLL population, and this is clearly an attractive area for future study, given the data for improved responses noted in healthy adult controls.

Pneumococcal Vaccines

A conjugated 13-valent pneumococcal vaccine called PCV13 (Prevnar 13; Wyeth/Pfizer) was FDA approved in 2010 for use in children and licensed for adults in 2011. Starting in 2012, ACIP pneumococcal vaccine guidelines recommended use of PCV13 along with the 23-valent pneumococcal polysaccharide vaccine (PPSV23) for immunocompromised adults aged 19 years and older, including those with congenital or acquired immunodeficiency, HIV infection, chronic kidney disease, nephrotic syndrome, leukemia, lymphoma, multiple myeloma, or other malignancy, and recipients of solid organ transplants.58 Limited data on the immunogenicity of PCV13 in immunocompromised adults are available.^{58,59} Similar or greater antibody responses to PCV13 compared with PPSV23 were found in immunocompetent adults.⁶⁰ According to ACIP guidelines, patients with CLL aged 19 years or older who have not previously received any pneumococcal vaccine should receive a dose of PCV13, followed by a dose of PPSV23 at least 8 weeks later (Table 3).58 For patients who have previously received 1 or more doses of PPSV23, 1 dose of PCV13 should be given at least 1 year after the last PPSV23 dose. According to ACIP, a second PPSV23 dose is recommended 5 years after the first PPSV23 dose for patients with CLL who are younger than 65 years.⁵⁸ For those who require additional doses of PPSV23, the first dose should be given at least 8 weeks after PCV13 and at least 5 years after the most recent dose of PPSV23 (see Table 3).58 No further doses of PPSV23 are needed for patients vaccinated with PPSV23 at or after age 65 years.⁵⁸

The schedule of pneumococcal vaccines (ie, order of conjugate and polysaccharide vaccines and interval between doses) has been shown to affect vaccine immunogenicity.60,61 Patients who receive PPSV23 followed 1 year later by PCV13 have decreased antibody responses compared with those who receive PCV13 followed 1 year later by PPSV23.62 These findings indicate that PCV13 augments the immune response to a subsequent dose of PPSV23 administered 1 year later for serotypes in common. Various study schedules of PCV13 and PPSV23 have confirmed that PCV13 is the preferred first choice for immunization of naive healthy adults to optimize antibody response, induction of memory, and maintenance of long-term protection. There have been concerns that repeated vaccination with pneumococcal polysaccharide vaccines may cause immune tolerance or vaccine hyporesponsiveness.⁶³

First Author	Study Type	Vaccine, Manufacturer; Dosing	Patient Data	Percent- age With Hypogamma- globulinemia (<6.2 g/L)	Percentage With Chemo- therapy/ Steroids	Immunogenicity Endpoint and Correlates
Van der Velden, ⁵⁶ 2001	Prospective cohort HI at days 0, 21, 42	Influvac (inactivated subunit) Duphar, Amsterdam, the Nether- lands 2 doses, 21 days apart	n=20 Rai 0, 20% Rai I/II, 40% Rai III/IV, 40%	15%	7 of 20 with previous chemother- apy, none in past 3 months	Seroconversion after 1 dose: 5% for H1N1 and H3N1, 15% for B. Seroconversion after 2 doses: 15% for H1N1 and H3N2, 30% for B. No significant increase in conversion or protection rates after booster. Influenza B: higher immuno- globulin levels were correlated with seroconversion.
Marotta, ⁵³ 1998	Prospective case-control HI at days 0, 30, 60	Isiflu Zonale (inactivated virus vaccine) Istituto Siero- vaccinogeno, Naples, Italy 2 doses, 31 days apart	n=18, CLL Binet A, 50% Binet B, 22% Binet C, 28% n=15, sex- and age-matched controls	22%	Not reported, no chemo- therapy in past 4 weeks	Did not assess seroconversion. Defined response as seropro- tection with HI titer >1:10. Correlation between response and absolute numbers of both CD4+/CD45RA+ naive T cells and CD5– B cells was found.
Bucalossi, ⁵⁵ 1995	Prospective case-control HI at days 0, 30, 60	Inflexal Berna (inactivated whole virus) Istituto Sieroterapico Berna, Como, Italy 2 doses, 31 days apart	n=30, CLL Binet A, 30% Binet B, 40% Binet C, 30% n=30, sex- and age-matched controls	Not reported	Not reported	Did not assess seroconversion. Defined response as seropro- tection with HI titer >1:10. Correlation between response and disease stage. Correlation between responses and immunoglobulin levels.

Table 2. Immunogenicity Studies of Influenza Vaccine in Patients With Chronic Lymphocytic Leukemia

HI, hemagglutinin inhibition assay.

Not all countries recommend repeated doses of PPSV23 for immunocompromised adults. Studies have shown that vaccine hyporesponsiveness does not seem to occur with repeated PPSV23 vaccination if more than 5 years have elapsed since the previous PPSV23 dose.⁶⁴⁻⁶⁶ Thus, ACIP recommends a period of 5 years or more between repeated PPSV23 doses (see Table 3).^{38,39}

Two studies have evaluated responses to pneumococcal vaccines in patients with CLL. One studied responses to the 23-valent polysaccharide pneumococcal vaccine, and the other studied responses to the 7-valent pneumococcal conjugate vaccine (Table 4). In the study of the polysaccharide vaccine, none of the 25 subjects had previously received the pneumococcal vaccine. The rate of response, defined as a 2-fold or greater increase in postvaccination serotype-specific IgG over prevaccination levels for at least 2 of 3 serotypes tested, was 22%. An adequate response to vaccination was seen only in patients with serum IgG levels in the normal range.⁶⁷ In the study of the 7-valent conjugate pneumococcal vaccine, none of the subjects had previously received either the polysaccharide or conjugate pneumococcal vaccines. In patients with CLL, the rate of vaccine response, defined as a 2-fold or greater increase in serotype-specific IgG and a postvaccination level of 0.35 µg/dL or higher for 6 of the 7 serotypes, was 24%, and the rate of vaccine response was 71% in controls (P<.001).68 The authors of this study noted that when the analysis was restricted to patients with Binet A disease, no prior chemotherapy, and no hypogammaglobulinemia, then 39% (11 of 28) of the patients with CLL responded to 6 serotypes, compared with only 5% (1 of 21) of the patients with CLL not meeting these 3 criteria (P<.007).⁶⁸ This

Influenza vaccines					
Age <65 y	Annual standard dose of inactivated influenza vaccine				
Age ≥65 y	Annual standard dose or high dose of inactivated influenza vaccine				
Tetanus-diphtheria, pertussis (Td, Tdap) vaccines					
No previous Tdap after age 11 y	Tdap regardless of interval since most recent Td vaccine and Td booster every 10 y thereafter				
Previous Tdap dose after age 11 y	Td booster every 10 y				
Pneumococcal vaccines					
PPSV23 vaccine history (number of previous doses)					
0	PCV13 first, followed by PPSV23	Second PPSV23 ^b	PPSV23 at 65 y ^b Adults who receive a dose of PPSV23 at age		
1	PCV13	Second PPSV23 ^b	≥65 y do not need another dose.		
2	PCV13				

Table 3. Recommendations for Vaccines Indicated in All Patients With Chronic Lymphocytic Leukemia ^a

Td, combined tetanus and diphtheria vaccine; Tdap, combined tetanus, diphtheria, and acellular pertussis vaccine; PPSV23, 23-valent pneumococcal polysaccharide vaccine; PCV13, conjugated 13-valent pneumococcal vaccine; w, weeks; y, years.

^a Indication for other vaccines depends on the patient's previous vaccination history, occupation, travel, and medical comorbidities.^{38,39}

^b Additional doses of PPSV23 should be administered 5 years or more after the previous PPSV23 dose if the patient is younger than 65 years of age.

would suggest that patients should be vaccinated early in the course of their disease and before the onset of hypogammaglobulinemia. However, the 39% response rate is still suboptimal for this patient group. Whether the new approach of PCV13 followed 8 weeks later by PPSV23 will improve immunogenicity in the CLL population merits evaluation.

Tetanus, Diphtheria, and Acellular Pertussis Vaccines

The tetanus-diphtheria (Td) vaccine should be administered every 10 years. According to Centers for Disease Control and Prevention ACIP recommendations, a single dose of acellular pertussis vaccine is recommended for previously unvaccinated persons aged 11 years or older.^{38,39} Therefore, all patients with CLL should receive a dose of tetanus-diphtheria-acellular pertussis (Tdap) vaccine if they have not yet received one in adulthood. Given the age of most patients with CLL, they would not have received the Tdap vaccine as adolescents because this vaccine is relatively new. The Tdap vaccine should be given even when the last Td vaccine was administered less than 10 years ago.^{38,39} Data on tetanus vaccine responses in CLL are sparse, and there are no data on responses to pertussis vaccines in CLL. One study noted decreased antibody responses to tetanus toxoid vaccine in patients with CLL compared with a control population. Higher rates of antibody responses to tetanus toxoid antigen were noted in those with early-stage disease and in those with normal serum Ig levels, a finding that would argue in favor updating Td vaccination shortly after diagnosis.69

Data regarding other vaccine responses in patients with CLL are scarce.⁷⁰ Responses to the tetanus toxoid–

Haemophilus influenzae type b conjugate vaccine have ranged from 21% to 48% of patients with CLL.^{67,69,71}

Vaccines to Avoid

In general, individuals with CLL should not receive any live attenuated viral or live attenuated bacterial vaccines (see Table 1). The contraindicated vaccines include intranasal influenza, measles, mumps, rubella, Sabin polio, bacille Calmette-Guérin (BCG), oral typhoid, varicella (chickenpox and zoster), and yellow fever vaccines. This recommendation is based upon the ACIP^{38,39} and IDSA⁴⁰ recommendations for immunocompromised persons. In a person with an inadequate immune system, there is the potential for a live vaccine to cause infection.

Strategies to Increase Vaccine Immunogenicity in Patients With Chronic Lymphocytic Leukemia

Based on the fact that plasma histamine levels have been observed to be higher in patients with CLL than in healthy controls,⁷¹ some have suggested that the production of antibodies against vaccines might be enhanced by blocking histamine type 2 receptors with ranitidine. Indeed, 2 small trials have suggested that ranitidine improves T-cell-dependent antibody responses to the *H influenzae* conjugate vaccine.^{71,72} However, ranitidine has not been successful in improving influenza vaccine⁷¹ or polysaccharide pneumococcal vaccine⁷² antibody responses. Although others have hypothesized that granulocyte-monocyte colony-stimulating factor (GM-CSF) might improve vaccine responses in patients with CLL, GM-CSF did not

First Author	Study Type	Vaccine, Manufacturer; Dosing	Patient Data	Percent- age With Hypogamma- globulinemia (<6.2 g/L)	Percentage With Che- motherapy/ Steroids	Immunogenicity Endpoint and Correlates
Hartkamp, ⁶⁷ 2001	Prospective Specific IgG titers against serotypes 3, 4, and 9 at days 0 and 21	Pneumovax 23; Merck Sharp & Dohme 1 dose	n=25, B-CLL Rai 0, 40% Rai I/II, 44% Rai III/IV, 16% Mean age, 70	Not reported	32% with previous che- motherapy, none with chemo- therapy 3 months before study	Response rate defined as ≥2-fold serotype-specific IgG increase for at least 2 of 3 serotypes tested; 22% responded. Response seen only in those with normal total IgG levels. Soluble CD23 levels were significantly lower in those with response.
Sinisalo, ⁶⁸ 2007	Prospective case-control Specific IgG titers against all 7 serotypes at days 0 and 21	Prevnar Pneumococ- cal 7-valent Conjugate Vaccine; Wyeth/ Pfizer 1 dose	n=49, CLL Binet A, 80% Binet B, 18% Binet C, 2% n=24, age- and sex-matched controls Mean age, 65	29%	22% with present or past chemo- therapy, 16% with ongoing steroids	Response rate defined as \geq 2-fold serotype-specific IgG increase + postvaccination level of \geq 0.35 µg/dL. Response to at least 6 serotypes: 24% in CLL patients and 71% in controls (<i>P</i> <.001). Statistically significant increase in response rate for controls vs CLL patients for all serotypes. If Binet A, before chemo- therapy, and no hypogamma- globulinemia, then 39% (11 of 28) of CLL patients responded to 6 serotypes, compared with only 5% (1 of 21) of other patients (<i>P</i> <.007).

Table 4. Immunogenicity Studies of Pneumococcal Vaccine in Patients With Chronic Lymphocytic Leukemia

IgG, immunoglobulin G; CLL, chronic lymphocytic leukemia.

improve polysaccharide pneumococcal vaccine antibody responses in controlled trials.⁷³ Lenalidomide has been shown to improve conjugate pneumococcal vaccine responses in patients with multiple myeloma.⁷⁴ Increases in serum Ig levels have been observed in patients treated with lenalidomide (Revlimid, Celgene).⁷⁵ Lenalidomide merits further testing as a vaccine adjuvant in patients with CLL.

New Directions

Strategies to increase vaccine immunogenicity in patients with CLL will likely not be a "one size fits all" approach. One plausible hypothesis is that some adjuvants will work for T-cell–dependent antibody responses and others will work for T-cell–independent antibody responses. In order to come up with rationally designed adjuvant strategies, we need to have a far better understanding of the mechanisms that lead to impaired vaccine responses in these patients. We propose that through detailed vaccinomics studies of vaccine responses in patients with CLL and in healthy age-matched controls, designed to uncover the exact mechanism(s) involved in deficient vaccine responses, we will be able to better elucidate the strategies that can be used to overcome these factors. Vaccinomics is the comprehensive study of immune responses to vaccines such that these responses can be understood, predicted, and then applied to the rational and directed development of vaccines.⁷⁶⁻⁷⁸ It relies upon the integration of the tools of transcriptomics, proteomics, epigenomics, immunogenetics, computational modeling, and immune monitoring.^{76,79} To a certain extent, our own studies on leukemic cell-stromal cell interactions have set the stage for such vaccinomics studies. In brief summary, there is significant evidence that leukemic B cells are profoundly immunosuppressive and modify stromal cell function to

favor disease progression.⁷⁴⁻⁷⁹ Vaccines have historically been developed based on an approach that included isolating a pathogen, inactivating or attenuating it, and then injecting it into a host. However, with a reliance upon the "-omics" tools listed above, which have allowed insights into individualized medicine, more informed vaccines and adjuvants can be designed to target subgroups of interest, such as immunosenescent populations, groups with common genetic variants that may impede the development of protective immune responses, and patients with malignant diseases, such as CLL, that cause profound immunosuppression.^{76,80-85}

The first steps in improving vaccine responses in CLL should include the detailed study of already-approved vaccines and schedules that have not yet been evaluated in this patient population: high-dose influenza vaccine, oilin-water adjuvant influenza vaccines currently licensed in Europe, and various combinations of the polysaccharide and conjugate pneumococcal vaccines that patients will be receiving as a result of the new pneumococcal vaccine guidelines. These studies should be designed so as to provide a clear understanding of the depth of vaccine responses in all stages of CLL and in the clinical MBL cohorts, including innate immune, adaptive humoral, cytokine, and cell-mediated immune responses.

With regard to MBL, no one has reported vaccine responses in MBL populations. As noted before, up to 3% to 12% of the healthy population may be found to have MBL. If individuals with these small, highly prevalent B-cell clones show suboptimal responses to vaccines, they may represent an important public health issue pertaining to deficient vaccine responses. If indeed individuals with MBL are not able to mount appropriate responses to vaccines, then alternative vaccine strategies, such as are administered to other immunocompromised groups, may need to be considered for use in this population.

Summary

Individuals with CLL appear to be at increased risk for infection, and evidence suggests that they have an inadequate response to most routinely used vaccines. Clinical trials are needed to address a number of key questions related to vaccine use in patients with CLL: (1) What are the seroconversion or seroprotection levels for all disease stages of CLL in response to specific vaccines? (2) What are the mechanisms of dysfunction related to suboptimal responses in CLL? (3) Do alternative vaccine preparations overcome the poor responses to certain pathogens? (4) Can vaccine adjuncts enhance responses to existing vaccines? (5) Can vaccinomics studies be employed in this patient population to elucidate and overcome the mechanisms responsible for decreased vaccine responses? Determining the responses to vaccines in patients with both low-count and high-count MBL will also be critically important and may have public health implications, given the prevalence of this condition. It is hoped that newer vaccines, alternative vaccination schedules, and the use of vaccine adjuvants may increase responses to vaccines in patients with CLL and reduce the number of deaths due to infection in this population of immunosuppressed patients.

Disclosures

Dr Whitaker receives funding for Mayo Clinic research from Pfizer Independent Grants for Learning and Change. Dr Shanafelt receives funding for Mayo Clinic research from Genentech, Celgene, Hospira, GlaxoSmithKline, Cephalon, and Janssen Pharmaceuticals. Dr Poland is chair of a safety evaluation committee for novel investigational vaccine trials being conducted by Merck Research Laboratories and offers consultative advice on vaccine development to Merck, CSL Biotherapies, Avianax, Sanofi Pasteur, Dynavax Technologies, Novartis Vaccines and Diagnostics, PaxVax, and Emergent Biosolutions, and he holds 2 patents related to vaccinia and measles peptide research. These activities have been reviewed by the Mayo Clinic Conflict of Interest Review Board and are conducted in compliance with Mayo Clinic conflict of interest policies. This research has been reviewed by the Mayo Clinic Conflict of Interest Review Board and was conducted in compliance with Mayo Clinic conflict of interest policies. Dr Kay receives research funding for Mayo Clinic research from Celgene, Hospira, Pharmacyclics, and Genentech, and he is on advisory or safety data monitoring boards for Celgene and Gilead.

References

1. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin.* 2011;61(2):69-90.

 Francis S, Karanth M, Pratt G, et al. The effect of immunoglobulin VH gene mutation status and other prognostic factors on the incidence of major infections in patients with chronic lymphocytic leukemia. *Cancer.* 2006;107(5):1023-1033.
 Shanafelt TD, Ghia P, Lanasa MC, Landgren O, Rawstron AC. Monoclonal B-cell lymphocytosis (MBL): biology, natural history and clinical management. *Leukemia.* 2010;24(3):512-520.

Shim YK, Middleton DC, Caporaso NE, et al. Prevalence of monoclonal B-cell lymphocytosis: a systematic review. *Cytometry B Clin Cytom.* 2010;78(suppl 1):S10-S18.
 Rawstron AC, Green MJ, Kuzmicki A, et al. Monoclonal B lymphocytes with the characteristics of "indolent" chronic lymphocytic leukemia are present in 3.5% of adults with normal blood counts. *Blood.* 2002;100(2):635-639.

6. Ghia P, Prato G, Scielzo C, et al. Monoclonal CD5+ and CD5- B-lymphocyte expansions are frequent in the peripheral blood of the elderly. *Blood.* 2004;103(6):2337-2342.
7. Nieto WG, Almeida J, Romero A, et al; Primary Health Care Group of Salamanca for the Study of MBL. Increased frequency (12%) of circulating chronic lymphocytic leukemia-like B-cell clones in healthy subjects using a highly sensitive multicolor flow cytometry approach. *Blood.* 2009;114(1):33-37.

Moreira J, Rabe KG, Cerhan JR, et al. Infectious complications among individuals with clinical monoclonal B-cell lymphocytosis (MBL): a cohort study of newly diagnosed cases compared to controls. *Leukemia*. 2013;27(1):136-141.
 Rawstron AC, Bennett FL, O'Connor SJ, et al. Monoclonal B-cell lymphocy-

 Rawstron AC, Bennett FL, O Connor SJ, et al. Monocional B-cell lymphocytosis and chronic lymphocytic leukemia. N Engl J Med. 2008;359(6):575-583. 10. Rawstron AC, Shanafelt T, Lanasa MC, et al. Different biology and clinical outcome according to the absolute numbers of clonal B-cells in monoclonal B-cell lymphocytosis (MBL). *Cytometry B Clin Cytom.* 2010;78(suppl 1):S19-S23.

11. Travade P, Dusart JD, Cavaroc M, Beytout J, Rey M. Severe infections associated with chronic lymphoid leukemia. 159 infectious episodes in 60 patients [in French]. *Presse Med.* 1986;15(34):1715-1718.

12. Tsiodras S, Samonis G, Keating MJ, Kontoyiannis DP. Infection and immunity in chronic lymphocytic leukemia. *Mayo Clin Proc.* 2000;75(10):1039-1054.

13. Hamblin AD, Hamblin TJ. The immunodeficiency of chronic lymphocytic leukaemia. *Br Med Bull.* 2008;87(1):49-62.

14. Kontoyiannis DP. Infection in chronic lymphocytic leukemia: a reappraisal. In: Cheson BD, ed. *Chronic Lymphocytic Leukemia: Scientific Advances and Clinical Developments*. New York, NY: Marcel Dekker; 1993:399-417.

15. Parikh S, Shanafelt TD, Rabe KG, et al. Hypogammaglobulinemia in patients with previously untreated chronic lymphocytic leukemia: clinical correlates and outcomes [ASH abstract 4178]. *Blood.* 2013;122(21).

16. Freeman JA, Crassini KR, Best OG, et al. Immunoglobulin G subclass deficiency and infection risk in 150 patients with chronic lymphocytic leukemia. *Leuk Lymphoma*. 2013;54(1):99-104.

17. Dearden C. Disease-specific complications of chronic lymphocytic leukemia. *Hematology Am Soc Hematol Educ Program.* 2008:450-456.

18. Morrison VA. Infectious complications of chronic lymphocytic leukaemia: pathogenesis, spectrum of infection, preventive approaches. *Best Pract Res Clin Haematol.* 2010;23(1):145-153.

19. Foon KA, Rai KR, Gale RP. Chronic lymphocytic leukemia: new insights into biology and therapy. *Ann Intern Med.* 1990;113(7):525-539.

20. Siegrist CA. Vaccine immunology. In: Plotkin SA, Orenstein WA, Offit PA, eds. *Vaccines*. 6th ed. Philadelphia, PA: Elsevier; 2013:14-32.

21. Molica S. Infections in chronic lymphocytic leukemia: risk factors, and impact on survival, and treatment. *Leuk Lymphoma*. 1994;13(3-4):203-214.

22. Chapel HM, Bunch C. Mechanisms of infection in chronic lymphocytic leukemia. *Semin Hematol.* 1987;24(4):291-296.

 Griffiths H, Lea J, Bunch C, Lee M, Chapel H. Predictors of infection in chronic lymphocytic leukaemia (CLL). *Clin Exp Immunol.* 1992;89(3):374-377.
 Scrivener S, Goddard RV, Kaminski ER, Prentice AG. Abnormal T-cell function

in B-cell chronic lymphocytic leukaemia. *Leuk Lymphoma*. 2003;44(3):383-389.
 Xay NE. Abnormal T-cell subpopulation function in CLL: excessive suppressor

(T gamma) and deficient helper (T mu) activity with respect to B-cell proliferation. *Blood.* 1981;57(3):418-420.

Platsoucas CD, Galinski M, Kempin S, Reich L, Clarkson B, Good RA. Abnormal T lymphocyte subpopulations in patients with B cell chronic lymphocytic leukemia: an analysis by monoclonal antibodies. *J Immunol.* 1982;129(5):2305-2312.
 Ziegler HW, Kay NE, Zarling JM. Deficiency of natural killer cell activity in patients with chronic lymphocytic leukemia. *Int J Cancer.* 1981;27(3):321-327.

28. Briggs PG, Kraft N, Atkins RC. T cells and CD45r expression in B-chronic lymphocytic leukemia. *Leuk Res.* 1990;14(2):155-159.

29. Ramsay AG, Johnson AJ, Lee AM, et al. Chronic lymphocytic leukemia T cells show impaired immunological synapse formation that can be reversed with an immunomodulating drug. *J Clin Invest.* 2008;118(7):2427-2437.

30. Goronzy JJ, Weyand CM. Understanding immunosenescence to improve responses to vaccines. *Nat Immunol*. 2013;14(5):428-436.

31. Lambert ND, Ovsyannikova IG, Pankratz VS, Jacobson RM, Poland GA. Understanding the immune response to seasonal influenza vaccination in older adults: a systems biology approach. *Expert Rev Vaccines*. 2012;11(8):985-994.

32. Oviedo-Orta E, Li CK, Rappuoli R. Perspectives on vaccine development for the elderly. *Curr Opin Immunol.* 2013;25(4):529-534.

33. Baluch A, Pasikhova Y. Influenza vaccination in oncology patients. *Curr Infect Dis Rep.* 2013;15(6):486-490.

34. Issa NC, Baden LR. Current issues in vaccines for adult patients with hematologic malignancies. *J Natl Compr Canc Netw.* 2012;10(11):1447-1454, quiz 1454.
35. Sempere L, Almenta I, Barrenengoa J, et al. Factors predicting response to hepatitis B vaccination in patients with inflammatory bowel disease. *Vaccine.* 2013;31(30):3065-3071.

36. Wallin L, Quintilio W, Locatelli F, Cassel A, Silva MB, Skare TL. Safety and efficiency of influenza vaccination in systemic lupus erythematosus patients. *Acta Reumatol Port*. 2009;34(3):498-502.

37. Guissa VR, Pereira RM, Sallum AM, et al. Influenza A H1N1/2009 vaccine in juvenile dermatomyositis: reduced immunogenicity in patients under immunosuppressive therapy. *Clin Exp Rheumatol.* 2012;30(4):583-588.

38. Bridges CB, Woods L, Coyne-Beasley T; ACIP Adult Immunization Work Group; Centers for Disease Control and Prevention (CDC). Advisory Committee on Immunization Practices (ACIP) recommended immunization schedule for adults aged 19 years and older--United States, 2013. *MMWR Surveill Summ*. 2013;62(suppl 1):9-19. 39. Bridges CB, Coyne-Beasley T; Advisory Committee on Immunization Practices (ACIP); ACIP Adult Immunization Work Group; Centers for Disease Control and Prevention (CDC). Advisory Committee on Immunization Practices recommended immunization schedule for adults aged 19 years or older - United States, 2014. *MMWR Morb Mortal Wkly Rep*. 2014;63(5):110-112.

40. Rubin LG, Levin MJ, Ljungman P, et al. 2013 IDSA clinical practice guideline for vaccination of the immunocompromised host. *Clin Infect Dis.* 2014;58(3):e44-e100. 41. Tsang P, Gorse GJ, Strout CB, et al. Immunogenicity and safety of Fluzone* intradermal and high-dose influenza vaccines in older adults ≥65 years of age: a randomized, controlled, phase II trial. *Vaccine.* 2014;32(21):2507-2517.

42. Sullivan SJ, Jacobson R, Poland GA. Advances in the vaccination of the elderly against influenza: role of a high-dose vaccine. *Expert Rev Vaccines*. 2010;9(10):1127-1133.

43. Fluzone high-dose vaccine is significantly more effective than the standard dose of fluzone vaccine in preventing influenza in adults ≥65 years of age. *Clin Infect Dis.* 2013;57(10):i, discussion i.

44. Roman F, Vaman T, Gerlach B, Markendorf A, Gillard P, Devaster JM. Immunogenicity and safety in adults of one dose of influenza A H1N1v 2009 vaccine formulated with and without AS03A-adjuvant: preliminary report of an observerblind, randomised trial. *Vaccine*. 2010;28(7):1740-1745.

45. McElhaney JE, Beran J, Devaster JM, et al; Influence65 study group. AS03adjuvanted versus non-adjuvanted inactivated trivalent influenza vaccine against seasonal influenza in elderly people: a phase 3 randomised trial. *Lancet Infect Dis.* 2013;13(6):485-496.

46. Durando P, Sticchi L, Alberti M, et al. Novel approaches for the prevention of influenza: the intradermal vaccination [in Italian]. Ig Sanita Pubbl. 2010;66(3):387-400.

 Mannino S, Villa M, Apolone G, et al. Effectiveness of adjuvanted influenza vaccination in elderly subjects in northern Italy. *Am J Epidemiol.* 2012;176(6):527-533.
 Gasparini R, Pozzi T, Montomoli E, et al. Increased immunogenicity of the MF59-adjuvanted influenza vaccine compared to a conventional subunit vaccine in elderly subjects. *Eur J Epidemiol.* 2001;17(2):135-140.

49. Podda A. The adjuvanted influenza vaccines with novel adjuvants: experience with the MF59-adjuvanted vaccine. *Vaccine*. 2001;19(17-19):2673-2680.

50. Ferguson M, Risi G, Davis M, et al. Safety and long-term humoral immune response in adults after vaccination with an H1N1 2009 pandemic influenza vaccine with or without AS03 adjuvant. *J Infect Dis.* 2012;205(5):733-744.

51. Cherif H, Höglund M, Pauksens K. Adjuvanted influenza a (H1N1) 2009 vaccine in patients with hematological diseases: good safety and immunogenicity even in chemotherapy-treated patients. *Eur J Haematol.* 2013;90(5):413-419.

52. Hottinger AF, George AC, Bel M, et al; H1N1 Study Group. A prospective study of the factors shaping antibody responses to the AS03-adjuvanted influenza A/H1N1 vaccine in cancer outpatients. *Oncologist.* 2012;17(3):436-445.

53. Marotta G, Bucalossi A, Galieni P, et al. CD4+/CD45RA+ 'naive' T cells and immunological response to influenza virus vaccine in B-cell chronic lymphocytic leukaemia patients. *Acta Haematol.* 1998;99(1):18-21.

54. Hannoun C, Megas F, Piercy J. Immunogenicity and protective efficacy of influenza vaccination. *Virus Res.* 2004;103(1-2):133-138.

55. Bucalossi A, Marotta G, Galieni P, Bigazzi C, Valenzin PE, Dispensa E. Immunological response to influenza virus vaccine in B-cell chronic lymphocytic leukaemia patients. *Acta Haematol.* 1995;93(1):56.

56. van der Velden AM, Mulder AH, Hartkamp A, Diepersloot RJ, van Velzen-Blad H, Biesma DH. Influenza virus vaccination and booster in B-cell chronic lymphocytic leukaemia patients. *Eur J Intern Med.* 2001;12(5):420-424.

57. Goodwin K, Viboud C, Simonsen L. Antibody response to influenza vaccination in the elderly: a quantitative review. *Vaccine*. 2006;24(8):1159-1169.

58. Centers for Disease Control and Prevention (CDC). Use of 13-valent pneumococcal conjugate vaccine and 23-valent pneumococcal polysaccharide vaccine for adults with immunocompromising conditions: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Morb Mortal Wkly Rep.* 2012;61(40):816-819.

 Smith KJ, Nowalk MP, Raymund M, Zimmerman RK. Cost-effectiveness of pneumococcal conjugate vaccination in immunocompromised adults. *Vaccine*. 2013;31(37):3950-3956.

60. Jackson LA, Gurtman A, Rice K, et al. Immunogenicity and safety of a 13-valent pneumococcal conjugate vaccine in adults 70 years of age and older previously vaccinated with 23-valent pneumococcal polysaccharide vaccine. *Vaccine*. 2013;31(35):3585-3593.

61. Centers for Disease Control and Prevention (CDC). Licensure of 13-valent pneumococcal conjugate vaccine for adults aged 50 years and older. *MMWR Morb Mortal Wkly Rep.* 2012;61(21):394-395.

62. Greenberg RN, Gurtman A, Frenck RW, et al. Sequential administration of 13-valent pneumococcal conjugate vaccine and 23-valent pneumococcal polysaccharide vaccine in pneumococcal vaccine-naïve adults 60-64 years of age. *Vaccine*. 2014;32(20):2364-2374.

63. O'Brien KL, Hochman M, Goldblatt D. Combined schedules of pneumococcal conjugate and polysaccharide vaccines: is hyporesponsiveness an issue? *Lancet Infect Dis.* 2007;7(9):597-606.

64. Hammitt LL, Bulkow LR, Singleton RJ, et al. Repeat revaccination with 23-valent pneumococcal polysaccharide vaccine among adults aged 55-74 years living in Alaska: no evidence of hyporesponsiveness. *Vaccine*. 2011;29(12):2287-2295. 65. Musher DM, Manoff SB, McFetridge RD, et al. Antibody persistence ten years after first and second doses of 23-valent pneumococcal polysaccharide vaccine, and immunogenicity and safety of second and third doses in older adults. *Hum Vaccin*. 2011;7(9):919-928.

66. Manoff SB, Liss C, Caulfield MJ, et al. Revaccination with a 23-valent pneumococcal polysaccharide vaccine induces elevated and persistent functional antibody responses in adults aged 65 > or = years. *J Infect Dis.* 2010;201(4):525-533.
67. Hartkamp A, Mulder AH, Rijkers GT, van Velzen-Blad H, Biesma DH. Antibody responses to pneumococcal and haemophilus vaccinations in patients with B-cell chronic lymphocytic leukaemia. *Vaccine.* 2001;19(13-14):1671-1677.

68. Sinisalo M, Vilpo J, Itälä M, Väkeväinen M, Taurio J, Aittoniemi J. Antibody response to 7-valent conjugated pneumococcal vaccine in patients with chronic lymphocytic leukaemia. *Vaccine*. 2007;26(1):82-87.

69. Sinisalo M, Aittoniemi J, Oivanen P, Käyhty H, Olander RM, Vilpo J. Response to vaccination against different types of antigens in patients with chronic lymphocytic leukaemia. *Br J Haematol.* 2001;114(1):107-110.

70. Sinisalo M, Aittoniemi J, Käyhty H, Vilpo J. Vaccination against infections in chronic lymphocytic leukemia. *Leuk Lymphoma.* 2003;44(4):649-652.

71. Jurlander J, de Nully Brown P, Skov PS, et al. Improved vaccination response during ranitidine treatment, and increased plasma histamine concentrations, in patients with B cell chronic lymphocytic leukemia. *Leukemia*. 1995;9(11):1902-1909.

72. Van der Velden AM, Van Velzen-Blad H, Claessen AM, et al. The effect of ranitidine on antibody responses to polysaccharide vaccines in patients with B-cell chronic lymphocytic leukaemia. *Eur J Haematol.* 2007;79(1):47-52.

73. Safdar A, Rodriguez GH, Rueda AM, et al. Multiple-dose granulocyte-mac-

rophage-colony-stimulating factor plus 23-valent polysaccharide pneumococcal vaccine in patients with chronic lymphocytic leukemia: a prospective, randomized trial of safety and immunogenicity. *Cancer.* 2008;113(2):383-387.

74. Noonan K, Rudraraju L, Ferguson A, et al. Lenalidomide-induced immunomodulation in multiple myeloma: impact on vaccines and antitumor responses. *Clin Cancer Res.* 2012;18(5):1426-1434.

75. Badoux XC, Keating MJ, Wen S, et al. Lenalidomide as initial therapy of elderly patients with chronic lymphocytic leukemia. *Blood.* 2011;118(13):3489-3498.

76. Poland GA, Kennedy RB, McKinney BA, et al. Vaccinomics, adversomics, and the immune response network theory: individualized vaccinology in the 21st century. *Semin Immunol.* 2013;25(2):89-103.

77. Poland GA, Ovsyannikova IG, Jacobson RM, Smith DI. Heterogeneity in vaccine immune response: the role of immunogenetics and the emerging field of vaccinomics. *Clin Pharmacol Ther.* 2007;82(6):653-664.

78. Ovsyannikova IG, Poland GA. Vaccinomics: current findings, challenges and novel approaches for vaccine development. *AAPS J.* 2011;13(3):438-444.

79. Poland GA, Ovsyannikova IG, Kennedy RB, Haralambieva IH, Jacobson RM. Vaccinomics and a new paradigm for the development of preventive vaccines against viral infections. *OMICS*. 2011;15(9):625-636.

80. Ding W, Knox TR, Tschumper RC, et al. Platelet-derived growth factor (PDGF)-PDGF receptor interaction activates bone marrow-derived mesenchymal stromal cells derived from chronic lymphocytic leukemia: implications for an angiogenic switch. *Blood.* 2010;116(16):2984-2993.

81. Ghosh AK, Secreto CR, Knox TR, Ding W, Mukhopadhyay D, Kay NE. Circulating microvesicles in B-cell chronic lymphocytic leukemia can stimulate marrow stromal cells: implications for disease progression. *Blood.* 2010;115(9):1755-1764.

82. Ding W, Nowakowski GS, Knox TR, et al. Bi-directional activation between mesenchymal stem cells and CLL B-cells: implication for CLL disease progression. *Br J Haematol.* 2009;147(4):471-483.

83. Zhang S, Kipps TJ. The pathogenesis of chronic lymphocytic leukemia. *Annu Rev Pathol.* 2014;9(1):103-118.

84. Kater AP, van Oers MH, Kipps TJ. Cellular immune therapy for chronic lymphocytic leukemia. *Blood.* 2007;110(8):2811-2818.

85. Cantwell M, Hua T, Pappas J, Kipps TJ. Acquired CD40-ligand deficiency in chronic lymphocytic leukemia. *Nat Med.* 1997;3(9):984-989.