Non-Clear Cell Renal Cell Carcinoma: Unpacking a Messy Term

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Keywords

Chromophobe renal cell carcinoma, molecularly defined renal cell carcinoma, non–clear cell renal cell carcinoma, papillary renal cell carcinoma, renal medullary carcinoma, 2022 World Health Organization Classification of Tumours of the Urinary System and Male Genital Organs Abstract: Non-clear cell renal cell carcinoma (nccRCC) is a heterogeneous category comprising approximately 25% of epithelial renal tumors. Unlike their far more common clear cell counterpart, many nccRCC subtypes are rare, poorly understood, and often aggressive in nature. Treatment options are very limited and generally have been unsuccessfully extrapolated from ccRCC trials. Numerous subtypes also seem to emerge in the context of inherited conditions or syndromes affecting younger individuals. In recent years, the World Health Organization classification of renal tumors has been frequently updated with the recognition of novel entities beyond ccRCC. The recommendations currently demonstrate a shift from morphology-based to molecularly based classification. Thus, aberrations in genes such as TFE3, TFEB, FH, SDH, SMARCB1, ELOC, and ALK define separate entities that cannot be distinguished on the basis of microscopic appearance alone. This review aims to deconstruct the general and nondescriptive term of nccRCC to provide a comprehensive presentation of specific subtypes, highlighting their epidemiologic associations and distinctive biological and clinicopathologic features. Delineating these complexities reveals areas that warrant improvement, which will translate into optimal diagnosis, treatment, and overall patient care.

Introduction

Approximately 75% of epithelioid renal carcinomas (RCCs) are categorized as clear cell RCC (ccRCC). $^{1\text{-}4}$ The remaining 25% are

Familial Syndrome or				RCC Risk or	
Disease Tuberous sclerosis	Inheritance AD	Genes TSC1, TSC2	RCC subtypes Chromophobe, eosinophilic solid and cystic RCC, hybrid chro- mophobe-oncocytoma, SDH-deficient RCC	Incidence 2%-4%	Other Associated Conditions Angiomyolipoma, subependymal giant cell astrocytoma, facial angiofibromas, renal cysts, eosino- philic vacuolated tumor, low-grade oncocytic tumor, others
PTEN hamartoma tumor syndrome	AD	PTEN	Clear cell RCC, papillary RCC, chromophobe RCC	34%	Breast cancer, endometrial cancer, thyroid cancer, colon cancer, gastrointestinal hamartomas, lipomas, Lhermitte-Duclos disease, macrocephaly (≥97th percentile), mucocutaneous lesions, others
Birt-Hogg-Dubé syndrome	AD	Folliculin	Chromophobe, hybrid chromophobe- oncocytoma, clear cell RCC, papillary (rare)	15%-25%	Skin fibrofolliculomas, oncocytoma, lung cysts, spontaneous pneumothorax
Hereditary leiomy- omatosis and RCC syndrome	AD	FH	Papillary type 2, FH-deficient RCC	21%	Cutaneous leiomyomas, uterine leiomyomas
Hereditary papillary renal cancer	AD	MET	Classic papillary RCC		Renal papillary adenomas
Pheochromocyto- ma-paraganglioma syndrome type 4	AD	SDH-B	SDH-deficient RCC	14%	Pheochromocytoma, paraganglioma, gastrointestinal stromal tumors, pulmonary chondroma, pituitary tumors
Sickle cell hemo- globinopathies/ trait	AR	HBB	Renal medullary carcinoma	Trait: 1/20,000	Sickle cell disease

Table. Non-Clear Cell Renal Cell Carcinoma Subtypes Associated With Hereditary Conditions

AD, autosomal dominant; AR, autosomal recessive; *FH*, fumarate hydratase; *HBB*, hemoglobin subunit beta; *PTEN*, phosphatase and tensin homolog; RCC, renal cell carcinoma; *SDH*, succinate dehydrogenase; *TSC*, tuberous sclerosis.

a histologically and molecularly heterogeneous group of tumor entities, commonly referred to as non-clear cell RCC (nccRCC), variant histology RCC, or divergent histology RCC.^{1,2} Named mainly in contradistinction to ccRCC because they generally lack prominent clear cell histology and VHL mutations, nccRCC is an umbrella term with minimal clinical relevance, given that it comprises numerous relatively rare cancer subtypes with distinct natural history, pathology, familial syndrome associations (Table), and treatment options. The presence of mixed phenotypes and the partial overlap in certain features among subtypes pose diagnostic challenges with serious implications for choice of treatment.^{5,6} This situation is further complicated by the scarcity of molecularly focused dedicated trials for each of these rare entities to inform evidence-based care recommendations.

Given the high variability of nccRCCs, further subclassification is warranted. Indeed, new entities have

continually been recognized over the last decades, thus raising the number of included RCC subtypes in the World Health Organization (WHO) classification of renal tumors from 12 in 2004 to 16 in 2016 and finally 21 in 2022.⁷ The most recent edition, the 5th, reveals a shift from morphologic to molecular classification and now employs driver genetic events to describe RCC entities regardless of their microscopic appearance (Figure).^{7,8}

This review aims to provide a comprehensive presentation of nccRCC subtypes and highlight their distinct clinicopathologic and basic molecular features. A brief overview of the current recommendations and trends in nccRCC management per subtype is included to inform research and aid clinicians in the differential diagnosis and treatment decisions for nccRCC subtypes. More detailed discussions of the therapeutic management considerations for nccRCC have been published elsewhere.^{1,2,9}

Morphology-Based Classification of nccRCC

nccRCC can be subclassified based on morphology alone into papillary RCC (pRCC), chromophobe RCC, collecting duct carcinoma (CDC), and other renal tumors.

Papillary RCC

pRCC, generally thought to originate from the proximal tubule epithelium,¹⁰ is the most common nccRCC histologic subtype, accounting for approximately 10% of all RCCs.⁴ pRCC demonstrates a strong male and Black race predominance.¹¹ For many years, pRCC was further histologically divided into type 1 and type 2. The 2022 WHO classification abolished the terms *type 1* and *type 2 pRCC*. Instead, it employs the term *classic papillary RCC* for the former type 1 pRCC and endorses further subcategorizations of the heterogeneous group of type 2 tumors according to their molecular features, such as *NF2*, fumarate hydratase (*FH*),^{12,13} and *ALK* alterations.^{7,8,14}

The classic pRCC variant was historically considered less aggressive and is commonly associated with MET alterations that may be therapeutically targeted. These alterations occur via either an increased chromosome 7 copy number or MET gene transcriptional upregulation, thus leading to pathway activation and the promotion of invasion and angiogenesis.¹⁵ In some cases, MET alterations are inherited in an autosomal dominant manner, and their carriers are at increased risk for the development of multiple, bilateral hereditary papillary renal carcinomas at a young age.^{16,17} An increased risk of the development of any pRCC variant has been associated with germline PTEN mutations in the context of PTEN hamartoma tumor syndrome/Cowden syndrome. Germline PTEN mutations are also associated with ccRCC and chromophobe RCC (Table).^{18,19}

pRCC is generally diagnosed in earlier stages and is on average less aggressive than ccRCC.^{20,21} However, it tends to be more resistant than ccRCC to immune checkpoint blockade and the targeted therapies developed primarily for ccRCC.^{1,2} The phase 2 PAPMET trial established cabozantinib (Cabometyx, Exelixis), a tyrosine kinase inhibitor (TKI) targeting multiple pathways, including vascular endothelial growth factor (VEGF) and MET, as a preferred targeted agent for metastatic pRCC.²² Cabozantinib can be used in combination with immunotherapy in aggressive disease when a rapid response is required.^{1,2} Lenvatinib (Lenvima, Eisai) plus everolimus is another treatment option than can yield responses in pRCC.²³ For what was formerly called type 2 pRCC, further classification based on mutational profile can reveal additional therapeutic targets. For example, molecular evaluation of tumors with papillary architectures can subclassify them as FH-deficient RCC or ALK-rearranged RCC, which

can be treated with bevacizumab plus erlotinib²⁴ or ALK inhibitors,¹⁴ respectively. These are further discussed in the section on molecularly defined RCC.

Chromophobe RCC

Chromophobe RCC, the third most common RCC subtype, accounts for 5% of RCC cases.⁴ It originates from the intercalated cells of the distal tubule.^{25,26} The 2 major putative mechanisms driving the pathogenesis of chromophobe RCC are mitochondrial dysfunction leading to oxidative stress and hyperactivation of the mechanistic target of rapamycin complex 1 (mTORC1) via PTEN pathway mutations.²⁶ The 2 main chromophobe RCC variants that have been described are classic and eosinophilic. Classic chromophobe RCC is characterized by a pale cytoplasm and irregular, hyperchromatic nuclei surrounded by a prominent cell membrane; the eosinophilic variant, as its name implies, is characterized by cells with purely or predominantly eosinophilic cytoplasm. On immunohistochemistry, positive staining for CD117 and CK7 in addition to PAX8 can aid in differentiating eosinophilic chromophobe RCC from more indolent tumors, such as oncocytomas.²⁷ Furthermore, chromophobe RCCs characteristically often show whole chromophobe copy number losses at chromosomes 1, 2, 6, 10, 13, 17, 21, and the sex chromosome while harboring a somatic mutation rate 3 times lower than that of ccRCC.²⁸ Chromophobe RCC is usually sporadic but can occur in the context of familial syndromes, often as multiple tumors.^{29,30} Birt-Hogg-Dubé syndrome, an autosomal dominant genodermatosis characterized by germline mutations in the folliculin (FLCN) gene, is known to cause RCCs, mostly of chromophobe or hybrid oncocytic-chromophobe histology, among other subtypes.^{29,31} PTEN hamartoma tumor syndrome (PTEN mutation) and tuberous sclerosis (TSC1 and TSC2 genes) also increase chromophobe RCC risk (Table).^{19,30}

Chromophobe RCC typically follows an indolent course and carries a better prognosis than ccRCC.²⁰ The prognosis appears to depend on tumor size and stage and the presence of histopathologic features such as sarcomatoid differentiation, vascular invasion, and microscopic necrosis.^{32,33} Whereas sarcomatoid dedifferentiation can occur in most RCC subtypes,³⁴ chromophobe RCC can also dedifferentiate into anaplastic, glandular, and neuroendocrine components often associated with more aggressive disease.^{35,36} Sarcomatoid dedifferentiation is associated with *TP53* and *PTEN* mutations followed by whole-genome duplication/imbalanced chromosomal duplication events, resulting in a flat copy number profile in comparison with the chromosomal losses found in conventional chromophobe RCC.^{28,36,37}

The currently available systemic therapies (immune checkpoint inhibitors and targeted agents) used for the

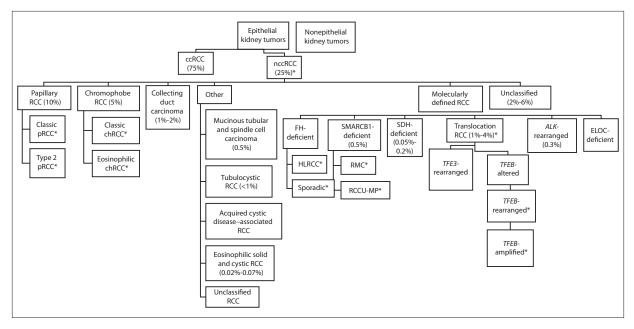


Figure. Renal cell carcinoma subtypes discussed in the text.

ALK, anaplastic lymphoma kinase; ccRCC, clear cell RCC; chRCC, chromophobe RCC; *ELOC*, elongin C; FH, fumarate hydratase; HLRCC, hereditary leiomyomatosis and RCC syndrome; nccRCC, non–clear cell RCC; pRCC, papillary RCC; RCC, renal cell carcinoma; RMC, renal medullary carcinoma. RCCU-MP, RCC unclassified with medullary phenotype; SDH, succinate dehydrogenase.

*Not a distinct entity in the 2022 World Health Organization classification.

treatment of ccRCC are far less effective against chromophobe RCC.^{1,2} Metastasis-directed locoregional therapies such as surgery, radiation therapy, and interventional radiology approaches are often prioritized when possible because conventional chromophobe RCC remains relatively indolent, even when metastatic.^{1,2,38} Although sarcomatoid dedifferentiation is found in only approximately 20% of patients with metastatic chromophobe RCC, it is associated with a highly aggressive polymetastatic course of disease, is refractory to currently available systemic therapies, and has a limited response to metastasis-directed locoregional therapies.^{1,2} Therefore, effective tailored treatment strategies for this RCC subtype are urgently needed.^{26,38}

Collecting Duct Carcinoma

CDC is a rare RCC subtype that arises from the collecting ducts and affects predominantly male patients.^{1,2,39} No distinct histologic patterns have been identified, but established diagnostic criteria require at least some involvement of the medullary region, a predominance of tubule formation, a desmoplastic stromal reaction, high-grade cytologic features, an infiltrative growth pattern, and the absence of other typical RCC subtypes or urothelial carcinoma.^{40,41} Nevertheless, CDC remains a diagnosis of exclusion and often morphologically overlaps with other malignancies, such as SMARCB1-deficient renal medullary carcinoma (RMC), FH-deficient RCC, and upper tract urothelial carcinoma.^{1,2,39-41}

CDC is often aggressive and is one of the few RCC subtypes that is sensitive to cytotoxic chemotherapy.^{1,2} Platinum-based chemotherapy, such as gemcitabine plus cisplatin or carboplatin plus paclitaxel, is the first-line option.^{1,2,42} The addition of bevacizumab may improve outcomes, but concerns over toxicity exist.⁴³ Among TKIs, cabozantinib is the best evaluated and produces a short progression-free survival of only 4 months.⁴⁴ Evidence regarding the efficacy of immune checkpoint therapy against CDC is limited to a few case reports.^{45,47}

Other Renal Tumors

Mucinous tubular and spindle cell carcinoma (MTSRCC) is a rare kidney tumor (~0.5% of RCCs) that affects predominantly female patients and is histologically characterized by tubular and spindle cell components on mucinous stroma.⁴⁸ *VSTM2A* overexpression, as determined by RNA in situ hybridization, is a sensitive and specific biomarker for MTSRCC.⁴⁹ MTSRCC is usually diagnosed at the localized stage, when it carries an excellent prognosis with nephrectomy alone. Advanced or metastatic cases have also been reported; these can respond to anti-VEGF TKIs and immune checkpoint therapy.⁵⁰⁻⁵² *CDKN2A/B* deletion and additional complex genomic abnormalities may contribute to the aggressive behavior of the rare metastatic cases of MTSRCC.⁵³ Hippo pathway deregulation resulting in increased levels of nuclear YAP1 protein is a recurrent hallmark of MTSRCC that may be therapeutically targeted.⁵⁴

Tubulocystic RCC is a rare entity that comprises fewer than 1% of RCCs and demonstrates a strong male predominance.⁵⁵ It has a distinctive histologic sponge-like appearance owing to the presence of multiple cysts filled with serous fluid, as well as a proliferation of tubules and a fibrous stroma.^{7,8,55,56} The most consistent molecular features include the loss of chromosomes 9 and Y, as well as gain of chromosome 17. Other chromosomal aberrations and mutations in chromatin-modifying genes can be present.7,8,55,56 Tubulocystic renal cell carcinoma is generally indolent, but occasional metastatic cases have been observed. Metastatic cases may morphologically resemble CDC, may harbor foci of sarcomatoid dedifferentiation, and can respond to anti-VEGF TKIs.56 Given the rarity of aggressive tubulocystic renal cell carcinoma, FH-deficient RCC should always be included in the differential diagnosis of metastatic RCCs with tubulocystic features.⁵⁷

Acquired cystic disease-associated renal cell carcinoma (ACD-RCC) is the neoplasm most strongly associated with end-stage renal disease, although end-stage renal disease (and the ensuing chronic inflammation) is a major risk factor for common RCC histologies, including ccRCC.⁵⁸ ACD-RCC affects predominantly male patients with a long history of dialysis and is often multiple or bilateral.⁵⁸ Histologically, ACD-RCC is described as cribriform or sieve-like with abundant granular eosinophilic cytoplasm. Hemorrhage, necrosis, and sarcomatoid histology were identified in higher-grade tumors.^{59,60} Chromosomes 3 and 16 aberrations and KMT2C and TSC2 mutations are the most common genetic features.58 ACD-RCC can occasionally be aggressive; 11% of patients experienced local or distant recurrence after surgery in a large case series of 40 patients.⁶⁰

Eosinophilic solid and cystic RCC (ESC RCC) is a novel entity in the 2022 classification that affects predominantly middle-aged women and accounts for fewer than 0.1% of RCC diagnoses.⁶¹ ESC RCC can reach large sizes and features solid and cystic areas of varying sizes. Cells are characterized by their eosinophilic cytoplasm and granular stippling.^{7,8,61} Its distinctive CK20 expression on immunohistochemistry can aid in the diagnosis.⁶¹ Most ESC tumors harbor somatic *TSC1* and *TSC2* gene mutations, whereas germline mutations—associated with tuberous sclerosis—are found in approximately 10% of patients.⁶¹⁻⁶³ The prognosis of ESC RCC is generally very favorable following treatment with radical or partial nephrectomy,⁶¹ although metastatic cases have occasionally been reported in the literature.^{62,64} The role of *TSC1* and *TSC2* mutations in ESC RCC provides a rationale for mTOR therapeutic inhibition. For example, a durable complete response to the mTOR inhibitor everolimus was noted following progression on 3 prior TKIs in a 13-yearold female patient with multifocal metastatic ESC RCC harboring a somatic *TSC2* mutation.⁶²

Molecularly Defined RCC

Molecularly defined RCC includes microphthalmia transcription factor (MiTF) family translocation RCC, FH-deficient RCC, SMARCB1-deficient RMC, succinate dehydrogenase (SDH)–deficient RCC, *ALK*-rearranged RCC, *ELOC*-mutated RCC, and unclassified RCC.

MiTF Family Translocation RCC

MiTF family translocation RCC accounts for approximately 1% to 2% of all sporadic RCC tumors and up to 50% of pediatric renal tumors, as well as approximately 15% of RCCs in adults younger than 45 years.⁵ The MiTF gene family consists of 4 members, among which TFE3 and TFEB have the best-established association with RCC.⁵ MiTF RCC tumors most commonly arise from oncogenic TFE3 rearrangements (translocations), followed by TFEB amplifications as the second most frequent cause and TFEB translocations as the third.^{5,65} The 2022 WHO RCC tumor classification removed the term MiTF translocation RCC, which is now further subclassified into TFE3-rearranged and TFEB-altered (TFEB-rearranged and TFEB-amplified) RCC because each of these subtypes has a very different biological behavior.7 Diagnosing TFE3-rearranged and TFEB-altered RCCs can be difficult because they may resemble ccRCC and pRCC on histologic examination.5,65 TFE3-rearranged RCC should be considered in children and young adults, particularly women, and in patients who have tumors with mixed morphologies-especially if they have a history of childhood chemotherapy; approximately 15% of TFE3-rearranged RCC cases are linked to childhood malignancies.⁶⁶

TFE3-rearranged RCC is also known as Xp11 translocation RCC on the basis of the *TFE3* gene locus on chromosome Xp11. Owing to its association with the X chromosome, *TFE3*-rearranged RCC most commonly arises in women: the female-to-male ratio is 2:1.⁶⁷ Multiple *TFE3* fusion partners have been identified; *ASPSCRI* (associated with alveolar soft part sarcomas), *PRCC*, and *SFPQ* are the most common and associated with a poor, an intermediate, and a favorable prognosis, respectively.⁵ Cell morphology on histology varies; papillary and nested patterns composed of clear or eosinophilic epithelioid cells are the most common, so that pRCC and ccRCC are involved in the differential diagnosis. Psammoma bodies are a common finding.⁶⁵ The diagnosis can be

made with TFE3 immunohistochemistry (IHC), breakapart fluorescence in situ hybridization (FISH), or RNA sequencing (RNA-seq).⁵ TFE3 IHC is highly sensitive but depends on the fixation processing and is less reliable than FISH or RNA-seq. TFE3 FISH is highly specific, but it cannot identify the TFE3 fusion partner and may result in false-negative results, particularly in cases due to cryptic intrachromosomal Xp11.2 inversions, including TFE3 fusions with the NONO or the RBM10 gene.⁵ RNA-seq is more costly but highly sensitive and can identify cryptic fusions and the exact partnering genes, which has important prognostic implications.⁵ Surgery is the appropriate strategy when the disease is confined to a kidney or regional lymph nodes. In more advanced settings, the immunotherapy and targeted agents used in other RCCs are commonly employed but are generally less effective than in ccRCC.^{1,2,68}

TFEB-altered RCC is less common and includes *TFEB*-rearranged and *TFEB*-amplified RCC. It often displays a nested histologic pattern with eosinophilic cells and can be diagnosed with IHC, *TFEB* break-apart FISH (which is different from *TFE3* FISH), or RNA-seq.^{5,65} *TFEB* is located on chromosome 6p21 and can be either amplified or rearranged to generate oncogenic fusions with an expanding list of potential partner genes, most commonly *MALAT1* on chromosome 11q12.⁶⁹ *TFEB*-amplified RCC typically affects older adults, with a slight male predominance, and is more common and aggressive than *TFEB*-rearranged RCC.⁶⁵ Anti-VEGF TKI treatment may be particularly effective against *TFEB*-amplified RCC owing to the frequent co-amplification of *VEGFA*, which is also mapped on chromosome 6p21.^{70,71}

FH-Deficient RCC

FH-deficient RCC typically occurs following somatic inactivation of the second FH allele in individuals with an autosomal dominant germline mutation in the other FH allele, a condition known as hereditary leiomyomatosis and RCC (HLRCC) syndrome. This syndrome is associated with FH-deficient RCC as well as cutaneous and uterine leiomyomas (Table).72,73 FH-deficient RCC may develop in up to 35% of individuals with HLRCC syndrome.⁷⁴ Sporadic cases are less common, comprising approximately 11% of all cases of FH-deficient RCC, and otherwise have pathologic features overlapping with those of the renal malignancies associated with germline HLRCC.^{6,75} Contrary to tumors related to other familial syndromes, FH-deficient RCC tumors in patients with HLRCC are usually solitary and unilateral but can occasionally be bilateral or multifocal.75 FH-deficient RCCs commonly demonstrate papillary architecture and were previously classified as type 2 pRCC tumors.^{7,8,75}

FH is a Krebs cycle enzyme catalyzing the conversion

of fumarate to malate.⁷⁶ FH inactivation leads to high fumarate levels, which result in the aberrant succination of proteins and the formation of products such as S-(2-succino)cysteine (2SC). Consequently, strong cytoplasmic and nuclear 2SC staining of tumor cells by IHC, with absent staining in the surrounding normal cells, is highly sensitive and specific for diagnosing FH-deficient RCC, even in rare cases of false-positive FH expression by IHC.⁷⁷

FH-deficient tumor cells depend on glycolysis for ATP production, and the increased fumarate levels stabilize HIF1A, leading to downstream VEGF transcription.⁷⁶ Accordingly, anti-VEGF TKIs and monoclonal antibodies such as bevacizumab can be effective therapies. The combination of bevacizumab and erlotinib achieved an overall response rate of 72.1% (95% CI, 57.2%-83.4%) and a median progression-free survival of 21.1 months (95% CI, 15.6-26.6) in patients with FH-deficient RCC.78 FH-deficient RCC frequently exhibits copy number gains on chromosome 7q, where MET is located, and retrospective data suggest that cabozantinib may be an effective treatment option for patients with FH-deficient RCC.^{1,2} Lenvatinib combined with either pembrolizumab (Keytruda, Merck) or everolimus can also induce responses in patients with FH-deficient RCC.79

SMARCB1-Deficient RMC

SMARCB1-deficient RMC most commonly affects individuals with a sickle hemoglobinopathy, such as sickle cell trait or sickle cell disease (Table).⁸⁰ SMARCB1, also known as INI1 or SNF5, is lost in RMC, most commonly owing to inactivating translocations or deletions that can be difficult to detect on standard clinical next-generation sequencing assays.⁸¹ Accordingly, the gold standard for RMC diagnosis is IHC showing loss of SMARCB1.⁸²

Owing to its association with sickle hemoglobinopathies, RMC most commonly occurs in young individuals of African descent, with a 2:1 male-to-female ratio.^{80,83} RMC is twice as likely to arise from the right kidney because the shorter vasculature of the left kidney is less susceptible to infarcts caused by red blood cell (RBC) sickling.^{80,84,85} Normal kidney cells downregulate SMARCB1 in response to hypoxia caused by RBC sickling in sickle cell trait, setting the stage for SMARCB1 loss and the eventual development of RMC.85 High-intensity exercise may further exacerbate RBC sickling, increasing RMC risk.⁸⁴ Hematuria is the most frequent presenting symptom, occurring in 60% of patients with RMC.83 Accordingly, RMC should always be suspected in young Black males with a history of high-intensity exercise who present with hematuria and a right-sided renal tumor. In such cases, hemoglobin electrophoresis should be performed if the sickle status is unknown, and a biopsy may be needed to confirm RMC up front. Chemotherapy rather than

nephrectomy is the recommended initial treatment, even in patients with RMC who present with radiologically nonmetastatic tumors greater than 4 cm in diameter.⁸² In up to 10% of cases, SMARCB1-deficient tumors with clinicopathologic features similar to those of RMC affect patients without sickle cell trait or a hemoglobinopathy.⁸³ These are known as RCC unclassified with medullary phenotype (RCCU-MP), have a slightly more favorable prognosis, and do not carry epidemiologic associations with male sex, right kidney laterality, and high-intensity exercise.⁸³

Platinum-based chemotherapy with carboplatin plus paclitaxel is the most commonly used first-line therapy for RMC.^{82,83} SMARCB1 loss induces DNA replication stress, sensitizing cells to DNA damage by platinum salts, nucleoside analogs such as gemcitabine, and topoisomerase inhibitors such as doxorubicin.81 Accordingly, the combination of gemcitabine with doxorubicin is an effective second-line strategy in RMC.86 Owing to upregulation of the epidermal growth factor receptor (EGFR) pathway in RMC,⁸¹ targeted therapies against EGFR are effective even in heavily pretreated patients.^{83,87} Combination definitive radiation therapy and chemotherapy can produce durable complete responses in selected patients with oligoprogressive or oligometastatic RMC.88 Earlier diagnosis and tailored therapeutic strategies in recent years have resulted in improved outcomes for patients with RMC.^{83,87} Elevated serum CA-125 levels are observed in two-thirds of patients with RMC and can be used to monitor therapeutic response.89

SDH-Deficient RCC

SDH participates both in the Krebs cycle, catalyzing the conversion of succinate to fumarate, and in the electron transport chain as part of the mitochondrial complex 2. SDH-deficient RCC is a rare renal malignancy, accounting for 0.05% to 0.2% of cases of RCC.90 It typically affects young and middle-aged patients, has a male predominance, and is associated with pheochromocytoma/ paraganglioma syndrome (Table), an autosomal dominant familial syndrome characterized by germline mutations of SDHA, SDHB, SDHC, SDHD, SDHAF2, MAX, or TMEM127. Patients with this syndrome are at risk for the development of pheochromocytoma, paraganglioma, gastrointestinal stromal tumors, and in rare cases pituitary tumors.⁹¹ Individuals with germline SDHB mutations have an increased lifetime risk (of approximately 5%) for the development of SDH-deficient RCC. The mean age at diagnosis is 38 years. The link between germline mutations in other SDH complex genes and SDH-deficient RCC has not been as firmly established.^{92,93} A total of 10 cases of SDHA-deficient RCC have been reported in the literature; SDHA and SDHB were negative by IHC, but patients had an uncertain germline status and a limited personal or family history of other SDH-related neoplasias.⁹⁴ In addition to *SDHA* mutations, 3 of these 10 tumors also harbored *NF2* gene mutations. Tumors with both *SDHA* and *NF2* mutations may show increased aggressiveness but respond well to immunotherapy.⁹⁴

The characteristic morphologic features of SDH-deficient RCC include eosinophilic cytoplasm with intracytoplasmic inclusions and vacuoles and indistinct cell borders.⁹⁰ SDHA-deficient RCCs typically lack this canonical morphology and instead demonstrate glandular, sheet-like, or papillary growth patterns with prominent nucleoli.⁹⁴ The diagnosis is established by loss of SDHB staining on IHC, regardless of the underlying mutated SDH subunit.^{90,93} In one study of 273 tumors originally diagnosed as oncocytomas, loss of SDHB with retention of SDHA by IHC was noted in 3 cases (1.1%), which were subsequently reclassified as SDH-deficient RCC.⁹⁵

Most SDH-deficient RCCs are low grade and have a favorable prognosis; long-term cure can be achieved with surgical resection alone. However, high-grade nuclear atypia, necrosis, and sarcomatoid dedifferentiation increase the risk of distant relapse, even decades after surgical resection of the primary SDH-deficient RCC, highlighting the need for long-term follow-up.^{90,93} Although additional data are required to determine effective treatment strategies for SDH-deficient RCC, the pseudo-hypoxic phenotype caused by SDH loss suggests that targeting the VEGF or HIF2A pathway is a reasonable approach.

ALK-Rearranged RCC

ALK-rearranged RCC is very rare and exhibits highly variable histology depending on the fusion partner gene. It can include tumors that would formerly have been classified as type 2 pRCC. The presence of a mucinous/ myxoid background or psammomatous calcifications can raise suspicion for ALK-rearranged RCC in tumors previously thought to be unclassified.⁹⁶ The ALK pathway is activated through fusion with various gene partners, with the EML4-ALK fusion demonstrating responsiveness to ALK inhibitors like alectinib (Alecensa, Genentech).¹⁴ Notably, the VCL-ALK fusion is found almost exclusively in individuals with sickle cell trait, but the tumor is far less aggressive than SMARCB1-deficient RMC.97 Almost all patients with ALK-rearranged RCC demonstrate ALK IHC positivity with a cytoplasmic and/or membranous distribution. The diagnosis can be confirmed with FISH or RNA-seq, with the latter providing the ability to identify the fusion partner.75

ELOC-Mutated RCC

Elongin C (ELOC; previously known as TCEB1) participates in the ubiquitination of VHL-bound HIF.

ELOC-mutated RCC was initially identified in 2013 in RCC with clear cell morphology that lacked VHL mutations or deletions in chromosome 3p, where VHL is located.98 Instead of ELOC mutations, some tumors may carry deletion of chromosome 8q, where ELOC is located.99 In addition to demonstrating clear cell histology without the VHL loss driving ccRCC, ELOC-mutated RCC is characterized by the presence of fibro-elastic bands or thick fibromuscular capsules and morphologic features overlapping with those of renal angiomyoadenomatous tumors, also known as RCC with (angio) leiomyomatous stroma. Renal angiomyoadenomatous tumors are distinct neoplasms associated with recurrent mutations in the TSC1/TSC2/mTOR pathway and show strong, diffuse glycoprotein nonmetastatic melanoma protein B (GPNMB) positivity on IHC, which is absent in ELOC-mutated RCC.^{100,101} A definitive diagnosis of ELOC-mutated RCC can be established by positive IHC staining for ELOC or molecular testing showing ELOC mutations.7,99 ELOC-mutated RCCs generally follow an indolent course, but the occasional metastatic cases can be treated with agents targeting the VEGF or HIF2A pathway.

Unclassified RCC

Unclassified RCC comprises a highly heterogeneous group of tumors that could not be classified morphologically or molecularly. Many of these cases are diagnosed at advanced stages, and the prognosis is variable.^{1,2} *NF2* loss resulting in Hippo pathway deregulation is the most common molecular event in unclassified RCC and is associated with a prognosis worse than that of tumors harboring other alterations, such as mTORC1 complex mutations.¹⁰² The growing recognition of new molecular and histomorphologic entities, along with the clinical application of comprehensive next-generation DNA and RNA sequencing, offer hope that fewer RCCs will fall under the unclassified category in the coming years.

Conclusion

There is a clear need to move beyond the term *nccRCC* and aim for more accurate diagnosis and classification. The development of dedicated clinical trials tailored to the specific clinical and molecular hallmarks of each RCC subtype can inform clinical practice.³⁸ A number of ongoing randomized clinical trials, such as PAPMET2, SAMETA, and STELLAR-304, represent a much needed push toward exploring therapeutic activity through randomized trials in nccRCC subtypes.⁹ Multicenter collaborations can help enroll well-selected patients, especially in the case of extremely rare nccRCC subtypes, for which randomization to a control arm may not be feasible.³⁸ These efforts

can be facilitated by the establishment of tissue biobanks for comprehensive molecular profiling, as well as cell line and animal models for functional preclinical studies. Search pages that can facilitate the identification of trials specific to selected histologies or molecular alterations are available from the Kidney Cancer Association (https:// www.kidneycancer.org/clinical-trials-finder) and KCCure (https://kccure.org/non-clear-cell-rcc-clinical-trials/).

Disclosures

Dr Msaouel has received honoraria for serving on the Scientific Advisory Boards of Mirati Therapeutics, BMS, and Exelixis; has done consulting for Axiom Healthcare Strategies; has participated in nonbranded educational programs supported by DAVA Oncology, Exelixis, and Pfizer; and has received research funding for clinical trials from Regeneron Pharmaceuticals, Summit Therapeutics, Takeda, BMS, Mirati Therapeutics, Gateway for Cancer Research, and the University of Texas MD Anderson Cancer Center.

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References

1. Msaouel P, Genovese G, Tannir NM. Renal cell carcinoma of variant histology: biology and therapies. *Hematol Oncol Clin North Am.* 2023;37(5):977-992.

2. Zoumpourlis P, Genovese G, Tannir NM, Msaouel P. Systemic therapies for the management of non-clear cell renal cell carcinoma: what works, what doesn't, and what the future holds. *Clin Genitourin Cancer*. 2021;19(2):103-116.

3. Alaghehbandan R, Siadat F, Trpkov K. What's new in the WHO 2022 classification of kidney tumours? *Pathologica*. 2022;115(1):8-22.

4. Padala SA, Barsouk A, Thandra KC, et al. Epidemiology of renal cell carcinoma. *World J Oncol.* 2020;11(3):79-87.

5. Tretiakova MS. Chameleon TFE3-translocation RCC and how gene partners can change morphology: accurate diagnosis using contemporary modalities. *Adv Anat Pathol.* 2022;29(3):131-140.

6. Chen YB. Update on selected high-grade renal cell carcinomas of the kidney: FH-deficient, ALK-rearranged, and medullary carcinomas. *Adv Anat Pathol.* 2024;31(2):118-125.

7. Goswami PR, Singh G, Patel T, Dave R. The WHO 2022 classification of renal neoplasms (5th edition): salient updates. *Cureus.* 2024;16(4):e58470.

8. Moch H, Amin MB, Berney DM, et al. The 2022 World Health Organization classification of tumours of the urinary system and male genital organs-part A: renal, penile, and testicular tumours. *Eur Urol.* 2022;82(5):458-468.

9. Salgia NJ, Zengin ZB, Pal SK, Dizman N. Renal cell carcinoma of variant histology: new biologic understanding leads to therapeutic advances. *Am Soc Clin Oncol Educ Book*. 2024;44(3):e438642.

10. Lindgren D, Sjölund J, Axelson H. Tracing renal cell carcinomas back to the nephron. *Trends Cancer.* 2018;4(7):472-484.

11. Sweeney PL, Jang A, Halat SK, Pal SK, Barata PC. Advanced papillary renal cell carcinoma: epidemiology, genomic drivers, current therapies, and ongoing trials. *Cancer Treat Res Commun.* 2022;33:100639.

12. Linehan WM, Spellman PT, Ricketts CJ, et al; Cancer Genome Atlas Research Network. Comprehensive molecular characterization of papillary renal-cell carcinoma. *N Engl J Med.* 2016;374(2):135-145.

13. Pal SK, Ali SM, Yakirevich E, et al. Characterization of clinical cases of advanced papillary renal cell carcinoma via comprehensive genomic profiling. *Eur Urol.* 2018;73(1):71-78.

14. Pal SK, Bergerot P, Dizman N, et al. Responses to alectinib in ALK-rearranged papillary renal cell carcinoma. *Eur Urol.* 2018;74(1):124-128.

15. Dizman N, Philip EJ, Pal SK. Genomic profiling in renal cell carcinoma. *Nat Rev Nephrol.* 2020;16(8):435-451.

16. Zbar B, Glenn G, Lubensky I, et al. Hereditary papillary renal cell carcinoma: clinical studies in 10 families. J Urol. 1995;153(3 pt 2):907-912.

17. Schmidt L, Duh FM, Chen F, et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. *Nat Genet*, 1997;16(1):68-73.

18. Lui ST, Shuch B. Genetic testing in kidney cancer patients: who, when, and how? *Eur Urol Focus.* 2019;5(6):973-976.

19. Mester JL, Zhou M, Prescott N, Eng C. Papillary renal cell carcinoma is associated with PTEN hamartoma tumor syndrome. *Urology*. 2012;79(5):1187.e1-7. 20. Abu-Ghanem Y, Powles T, Capitanio U, et al. The impact of histological subtype on the incidence, timing, and patterns of recurrence in patients with renal cell carcinoma after surgery-results from RECUR consortium. *Eur Urol Oncol*. 2021;4(3):473-482.

21. Steffens S, Janssen M, Roos FC, et al. Incidence and long-term prognosis of papillary compared to clear cell renal cell carcinoma—a multicentre study. *Eur J Cancer.* 2012;48(15):2347-2352.

22. Pal SK, Tangen C, Thompson IM Jr, et al. A comparison of sunitinib with cabozantinib, crizotinib, and savolitinib for treatment of advanced papillary renal cell carcinoma: a randomised, open-label, phase 2 trial. *Lancet.* 2021;397(10275):695-703.

23. Hutson TE, Michaelson MD, Kuzel TM, et al. A single-arm, multicenter, phase 2 study of lenvatinib plus everolimus in patients with advanced non-clear cell renal cell carcinoma. *Eur Urol.* 2021;80(2):162-170.

24. Choi Y, Keam B, Kim M, et al. Bevacizumab plus erlotinib combination therapy for advanced hereditary leiomyomatosis and renal cell carcinoma-associated renal cell carcinoma: a multicenter retrospective analysis in Korean patients. *Cancer Res Treat.* 2019;51(4):1549-1556.

25. Störkel S, Steart PV, Drenckhahn D, Thoenes W. The human chromophobe cell renal carcinoma: its probable relation to intercalated cells of the collecting duct. *Virchows Arch B Cell Pathol Incl Mol Pathol.* 1989;56(4):237-245.

26. Henske EP, Cheng L, Hakimi AA, Choueiri TK, Braun DA. Chromophobe renal cell carcinoma. *Cancer Cell.* 2023;41(8):1383-1388.

27. Alaghehbandan R, Przybycin CG, Verkarre V, Mehra R. Chromophobe renal cell carcinoma: novel molecular insights and clinicopathologic updates. *Asian J Urol.* 2022;9(1):1-11.

28. Lobo J, Ohashi R, Amin MB, et al. WHO 2022 landscape of papillary and chromophobe renal cell carcinoma. *Histopathology*. 2022;81(4):426-438.

29. Benusiglio PR, Giraud S, Deveaux S, et al; French National Cancer Institute Inherited Predisposition to Kidney Cancer Network. Renal cell tumour characteristics in patients with the Birt-Hogg-Dubé cancer susceptibility syndrome: a retrospective, multicentre study. *Orphanet J Rare Dis.* 2014;9(1):163.

30. Guo J, Tretiakova MS, Troxell ML, et al. Tuberous sclerosis-associated renal cell carcinoma: a clinicopathologic study of 57 separate carcinomas in 18 patients. *Am J Surg Pathol.* 2014;38(11):1457-1467.

31. Pavlovich CP, Walther MM, Eyler RA, et al. Renal tumors in the Birt-Hogg-Dubé syndrome. *Am J Surg Pathol.* 2002;26(12):1542-1552.

32. Casuscelli J, Becerra MF, Seier K, et al. Chromophobe renal cell carcinoma: results from a large single-institution series. *Clin Genitourin Cancer*. 2019;17(5):373-379.e4.

33. Przybycin CG, Cronin AM, Darvishian F, et al. Chromophobe renal cell car-

cinoma: a clinicopathologic study of 203 tumors in 200 patients with primary resection at a single institution. *Am J Surg Pathol.* 2011;35(7):962-970.

34. Hahn AW, Lebenthal J, Genovese G, Sircar K, Tannir NM, Msaouel P. The significance of sarcomatoid and rhabdoid dedifferentiation in renal cell carcinoma. *Cancer Treat Res Commun.* 2022;33:100640.

35. Alaghehbandan R, Williamson SR, McKenney JK, Hes O. The histologic diversity of chromophobe renal cell carcinoma with emphasis on challenges encountered in daily practice. *Adv Anat Pathol.* 2022;29(4):194-207.

36. Kapur P, Zhong H, Le D, et al. Molecular underpinnings of dedifferentiation and aggressiveness in chromophobe renal cell carcinoma. *JCI Insight*. 2024;9(10):e176743.

37. Collins K, Acosta AM, Siegmund SE, Cheng L, Hirsch MS, Idrees MT. Genetic profiling uncovers genome-wide loss of heterozygosity and provides insight into mechanisms of sarcomatoid transformation in chromophobe renal cell carcinoma. *Mod Pathol.* 2024;37(2):100396.

38. Msaouel P, Sheth RA. Locoregional therapies in immunologically "cold" tumors: opportunities and clinical trial design considerations. *J Vasc Interv Radiol.* 2024;35(2):198-202.

39. Sui W, Matulay JT, Robins DJ, et al. Collecting duct carcinoma of the kidney: disease characteristics and treatment outcomes from the National Cancer Database. *Urol Oncol.* 2017;35(9):540.e13-540.e18.

40. Cabanillas G, Montoya-Cerrillo D, Kryvenko ON, Pal SK, Arias-Stella JA III. Collecting duct carcinoma of the kidney: diagnosis and implications for management. *Urol Oncol.* 2022;40(12):525-536.

41. Singh JA, Ohe C, Smith SC. High grade infiltrative adenocarcinomas of renal cell origin: new insights into classification, morphology, and molecular pathogenesis. *Pathol Int.* 2018;68(5):265-277.

42. Oudard S, Banu E, Vieillefond A, et al; GETUG (Groupe d'Etudes des Tumeurs Uro-Génitales). Prospective multicenter phase II study of gemcitabine plus platinum salt for metastatic collecting duct carcinoma: results of a GETUG (Groupe d'Etudes des Tumeurs Uro-Génitales) study. *J Urol.* 2007;177(5):1698-1702.

43. Pécuchet N, Bigot F, Gachet J, et al. Triple combination of bevacizumab, gemcitabine and platinum salt in metastatic collecting duct carcinoma. *Ann Oncol.* 2013;24(12):2963-2967.

44. Procopio G, Sepe P, Claps M, et al. Cabozantinib as first-line treatment in patients with metastatic collecting duct renal cell carcinoma: results of the BON-SAI trial for the Italian Network for Research in Urologic-Oncology (Meet-URO 2 study). *JAMA Oncol.* 2022;8(6):910-913.

45. Danno T, Iwata S, Niimi F, Honda S, Okada H, Azuma T. Nivolumab and ipilimumab combination immunotherapy for patients with metastatic collecting duct carcinoma. *Case Rep Urol.* 2021;2021:9936330.

46. Yasuoka S, Hamasaki T, Kuribayashi E, et al. Nivolumab therapy for metastatic collecting duct carcinoma after nephrectomy: a case report. *Medicine (Baltimore)*. 2018;97(45):e13173.

47. Pyrgidis N, Sokolakis I, Haltmair G, Heller V, Hatzichristodoulou G. Avelumab in metastatic collecting duct carcinoma of the kidney: a case report. *J Med Case Rep.* 2023;17(1):262.

48. Xu X, Zhong J, Zhou X, et al. Mucinous tubular and spindle cell carcinoma of the kidney: a study of clinical, imaging features and treatment outcomes. *Front Oncol.* 2022;12:865263.

49. Kwon R, Argani P, Epstein JI, et al. Contemporary characterization and recategorization of adult unclassified renal cell carcinoma. *Am J Surg Pathol.* 2021;45(4):450-462.

50. Ling C, Tan R, Li J, Feng J. Mucinous tubular and spindle cell carcinoma of the kidney: a report of seven cases. *BMC Cancer.* 2023;23(1):815.

51. Fuchizawa H, Kijima T, Takada-Owada A, et al. Metastatic mucinous tubular and spindle cell carcinoma of the kidney responding to nivolumab plus ipilimumab. *IJU Case Rep.* 2021;4(5):333-337.

52. Ged Y, Chen YB, Knezevic A, et al. Mucinous tubular and spindle-cell carcinoma of the kidney: clinical features, genomic profiles, and treatment outcomes. *Clin Genitourin Cancer.* 2019;17(4):268-274.e1.

53. Yang C, Cimera RS, Aryeequaye R, et al. Adverse histology, homozygous loss of CDKN2A/B, and complex genomic alterations in locally advanced/metastatic renal mucinous tubular and spindle cell carcinoma. *Mod Pathol.* 2021;34(2):445-456.

54. Mehra R, Vats P, Cieslik M, et al. Biallelic alteration and dysregulation of the Hippo pathway in mucinous tubular and spindle cell carcinoma of the kidney. *Cancer Discov.* 2016;6(11):1258-1266.

55. Bhullar JS, Varshney N, Bhullar AK, Mittal VK. A new type of renal cancer tubulocystic carcinoma of the kidney: a review of the literature. *Int J Surg Pathol.* 2014;22(4):297-302.

56. Delahunt B, Srigley JR. The evolving classification of renal cell neoplasia. *Semin Diagn Pathol.* 2015;32(2):90-102.

57. Smith SC, Trpkov K, Chen YB, et al. Tubulocystic carcinoma of the kidney with poorly differentiated foci: a frequent morphologic pattern of fumarate hydratase-deficient renal cell carcinoma. *Am J Surg Pathol.* 2016;40(11):1457-1472.

58. Duong NX, Le MK, Nguyen TT, et al. Acquired cystic disease-associated renal cell carcinoma: a systematic review and meta-analysis. *Clin Genitourin Cancer.* 2024;22(3):102050.

59. Kondo T, Sasa N, Yamada H, et al. Acquired cystic disease-associated renal cell carcinoma is the most common subtype in long-term dialyzed patients: central pathology results according to the 2016 WHO classification in a multi-institu-tional study. *Pathol Int.* 2018;68(10):543-549.

60. Przybycin CG, Harper HL, Reynolds JP, et al. Acquired cystic disease-associated renal cell carcinoma (ACD-RCC): a multiinstitutional study of 40 cases with clinical follow-up. *Am J Surg Pathol.* 2018;42(9):1156-1165.

61. Trpkov K, Hes O, Bonert M, et al. Eosinophilic, solid, and cystic renal cell carcinoma: clinicopathologic study of 16 unique, sporadic neoplasms occurring in women. *Am J Surg Pathol.* 2016;40(1):60-71.

62. Palsgrove DN, Li Y, Pratilas CA, et al. Eosinophilic solid and cystic (ESC) renal cell carcinomas harbor TSC mutations: molecular analysis supports an expanding clinicopathologic spectrum. *Am J Surg Pathol.* 2018;42(9):1166-1181.

63. Mehra R, Vats P, Cao X, et al. Somatic bi-allelic loss of TSC genes in eosinophilic solid and cystic renal cell carcinoma. *Eur Urol.* 2018;74(4):483-486.

64. Tretiakova MS. Eosinophilic solid and cystic renal cell carcinoma mimicking epithelioid angiomyolipoma: series of 4 primary tumors and 2 metastases. *Hum Pathol.* 2018;80:65-75.

65. Skala SL, Xiao H, Udager AM, et al. Detection of 6 TFEB-amplified renal cell carcinomas and 25 renal cell carcinomas with MITF translocations: systematic morphologic analysis of 85 cases evaluated by clinical TFE3 and TFEB FISH assays. *Mod Pathol*, 2018;31(1):179-197.

66. Argani P, Laé M, Ballard ET, et al. Translocation carcinomas of the kidney after chemotherapy in childhood. *J Clin Oncol.* 2006;24(10):1529-1534.

67. Achom M, Sadagopan A, Bao C, et al. A genetic basis for sex differences in Xp11 translocation renal cell carcinoma. *Cell.* 2024;187(20):5735-5752.e25.

68. Alhalabi O, Thouvenin J, Négrier S, et al. Immune checkpoint therapy combinations in adult advanced MiT family translocation renal cell carcinomas. *Oncologist.* 2023;28(5):433-439.

69. Qu Y, Wu X, Anwaier A, et al. Proteogenomic characterization of MiT family translocation renal cell carcinoma. *Nat Commun.* 2022;13(1):7494.

70. Gupta S, Johnson SH, Vasmatzis G, et al. TFEB-VEGFA (6p21.1) co-amplified renal cell carcinoma: a distinct entity with potential implications for clinical management. *Mod Pathol.* 2017;30(7):998-1012.

71. Takamori H, Maeshima AM, Kato I, Baba M, Nakamura E, Matsui Y. TFEB-translocated and -amplified renal cell carcinoma with VEGFA co-amplification: a case of long-term control by multimodal therapy including a vascular endothelial growth factor-receptor inhibitor. *IJU Case Rep.* 2023;6(3):161-164.

72. Trpkov K, Hes O, Agaimy A, et al. Fumarate hydratase-deficient renal cell carcinoma is strongly correlated with fumarate hydratase mutation and hereditary leiomyomatosis and renal cell carcinoma syndrome. *Am J Surg Pathol.* 2016;40(7):865-875.

73. Carter CS, Skala SL, Chinnaiyan AM, et al. Immunohistochemical characterization of fumarate hydratase (FH) and succinate dehydrogenase (SDH) in cutaneous leiomyomas for detection of familial cancer syndromes. *Am J Surg Pathol.* 2017;41(6):801-809.

74. Skala SL, Dhanasekaran SM, Mehra R. Hereditary leiomyomatosis and renal cell carcinoma syndrome (HLRCC): a contemporary review and practical discussion of the differential diagnosis for HLRCC-associated renal cell carcinoma. *Arch Pathol Lab Med.* 2018;142(10):1202-1215.

75. Baniak N, Tsai H, Hirsch MS. The differential diagnosis of medullary-based renal masses. *Arch Pathol Lab Med.* 2021;145(9):1148-1170.

76. Linehan WM, Rouault TA. Molecular pathways: fumarate hydratase-deficient kidney cancer—targeting the Warburg effect in cancer. *Clin Cancer Res.* 2013;19(13):3345-3352.

77. Mannan R, Wang X, Bawa PS, et al. Characterization of protein S-(2-succino)-cysteine (2SC) succination as a biomarker for fumarate hydratase-deficient renal cell carcinoma. *Hum Pathol.* 2023;134:102-113.

78. Srinivasan R, Gurram S, Al Harthy M, et al. Results from a phase II study of bevacizumab and erlotinib in subjects with advanced hereditary leiomyomatosis and renal cell cancer (HLRCC) or sporadic papillary renal cell cancer [ASCO abstract 5004]. *J Clin Oncol.* 2020;38(15)(suppl).

79. Gleeson JP, Nikolovski I, Dinatale R, et al. Comprehensive molecular characterization and response to therapy in fumarate hydratase-deficient renal cell carcinoma. *Clin Cancer Res.* 2021;27(10):2910-2919.

80. Msaouel P, Tannir NM, Walker CL. A model linking sickle cell hemoglobinopathies and SMARCB1 loss in renal medullary carcinoma. *Clin Cancer Res.* 2018;24(9):2044-2049.

81. Msaouel P, Malouf GG, Su X, et al. Comprehensive molecular characterization identifies distinct genomic and immune hallmarks of renal medullary carcinoma. *Cancer Cell.* 2020;37(5):720-734.e13.

82. Msaouel P, Hong AL, Mullen EA, et al. Updated recommendations on the diagnosis, management, and clinical trial eligibility criteria for patients with renal medullary carcinoma. *Clin Genitourin Cancer.* 2019;17(1):1-6.

83. Lebenthal JM, Kontoyiannis PD, Hahn AW, et al. Clinical characteristics, management, and outcomes of patients with renal medullary carcinoma: a single-center retrospective analysis of 135 patients. *Eur Urol Oncol.* 2024;S2588-9311(24)00175-5.

84. Shapiro DD, Soeung M, Perelli L, et al. Association of high-intensity exercise with renal medullary carcinoma in individuals with sickle cell trait: clinical observations and experimental animal studies. *Cancers (Basel)*. 2021;13(23):6022.
85. Soeung M, Perelli L, Chen Z, et al. SMARCB1 regulates the hypoxic stress response in sickle cell trait. *Proc Natl Acad Sci USA*. 2023;120(21):e2209639120.
86. Wilson NR, Wiele AJ, Surasi DS, et al. Efficacy and safety of gemcitabine plus doxorubicin in patients with renal medullary carcinoma. *Clin Genitourin Cancer*. 2021;19(6):e401-e408.

Wiele AJ, Surasi DS, Rao P, et al. Efficacy and safety of bevacizumab plus erlotinib in patients with renal medullary carcinoma. *Cancers (Basel)*. 2021;13(9):2170.
 Mbilinyi RH, Msaouel P, Rao P, Karam JA, Tannir NM, Tang C. Radiation therapy for the management of renal medullary carcinoma: a multi-case study. *Clin Genitourin Cancer*. 2024;22(3):102065.

89. Grimm SL, Karki M, Blum KA, et al. CA-125 as a biomarker in renal medullary carcinoma: integrated molecular profiling, functional characterization, and prospective clinical validation [published online January 21, 2025]. *Clin Cancer Res.* doi:10.1158/1078-0432.CCR-24-3324.

90. Gill AJ, Hes O, Papathomas T, et al. Succinate dehydrogenase (SDH)-deficient renal carcinoma: a morphologically distinct entity: a clinicopathologic series of 36 tumors from 27 patients. *Am J Surg Pathol.* 2014;38(12):1588-1602.

91. Rogala J, Zhou M. Hereditary succinate dehydrogenase-deficient renal cell carcinoma. *Semin Diagn Pathol.* 2024;41(1):32-41.

92. Carlo MI. Hereditary renal cell carcinoma syndromes. *Hematol Oncol Clin North Am.* 2023;37(5):841-848.

93. Moch H, Ohashi R, Gandhi JS, Amin MB. Morphological clues to the appropriate recognition of hereditary renal neoplasms. *Semin Diagn Pathol.* 2018;35(3):184-192.

94. Liu J, Wang Y, Wang X, et al. Succinate dehydrogenase A deficient renal cell carcinoma: a rare renal tumor distinct from typical succinate dehydrogenase deficient renal cell carcinoma. *Pathol Res Pract.* 2024;261:155459.

95. Gupta S, Swanson AA, Chen YB, et al. Incidence of succinate dehydrogenase and fumarate hydratase-deficient renal cell carcinoma based on immunohistochemical screening with SDHA/SDHB and FH/2SC. *Hum Pathol.* 2019;91:114-122.

96. Trpkov K, Williamson SR, Gill AJ, et al. Novel, emerging and provisional renal entities: the Genitourinary Pathology Society (GUPS) update on renal neoplasia. *Mod Pathol.* 2021;34(6):1167-1184.

97. Wangsiricharoen S, Zhong M, Ranganathan S, Matoso A, Argani P. ALK-rearranged renal cell carcinoma (RCC): a report of 2 cases and review of the literature emphasizing the distinction between VCL-ALK and non-VCL-ALK RCC. *Int J Surg Pathol.* 2021;29(7):808-814.

98. Sato Y, Yoshizato T, Shiraishi Y, et al. Integrated molecular analysis of clear-cell renal cell carcinoma. *Nat Genet.* 2013;45(8):860-867.

99. Wang Y, Zhao P, Wang L, et al. Analysis of clinicopathological and molecular features of ELOC(TCEB1)-mutant renal cell carcinoma. *Pathol Res Pract.* 2022;235:153960.

100. Shah RB, Stohr BA, Tu ZJ, et al. "Renal cell carcinoma with leiomyomatous stroma" harbor somatic mutations of TSC1, TSC2, MTOR, and/or ELOC (TCEB1): clinicopathologic and molecular characterization of 18 sporadic tumors supports a distinct entity. *Am J Surg Pathol.* 2020;44(5):571-581.

101. Shah RB, Mehra R. Renal cell carcinoma associated with TSC/MTOR genomic alterations: an update on its expanding spectrum and an approach to clinicopathologic work-up. *Adv Anat Pathol.* 2024;31(2):105-117.

102. Chen YB, Xu J, Skanderup AJ, et al. Molecular analysis of aggressive renal cell carcinoma with unclassified histology reveals distinct subsets. *Nat Commun.* 2016;7(1):13131.